



# **Myxosporeans and myxosporidiosis of common carp and gibel carp in China**

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Hungary  
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# Brief history of fish myxosporean research in China

## IV developmental stages

**I. Initial stage:** 1949-1959;  
description solely based on morphology of myxospores

**II. Developmental stage:** 1960-1998;  
traditional taxonomy, epidemiology, histopathology  
Hallmark work: Publication of *Fauna Sinica, Myxozoa, Myxosporea* in 1998

**III. Mature stage:** 1998-2013;  
combined morphological and molecular characteristics, host, tissue tropism; molecular epidemiology; life cycle, ecology, host-parasite relationships;

**IV. Progressive stage:** 2013 afterwards;  
Genomic era; evolution; phylogenomic analysis; functional genomics  
Hallmark work: available of full genome data of *Thelohanellus kitauei*; ongoing genomic project of *Myxobolus honghuensis*



Prof. Chen



Prof. Ma



Recorded 575  
species in 23  
genera 1998



2.7 million tons/year;  
third among freshwater  
cultured fish;  
About 70 myxosporean  
species reported

Most only with  
simple morphological  
description

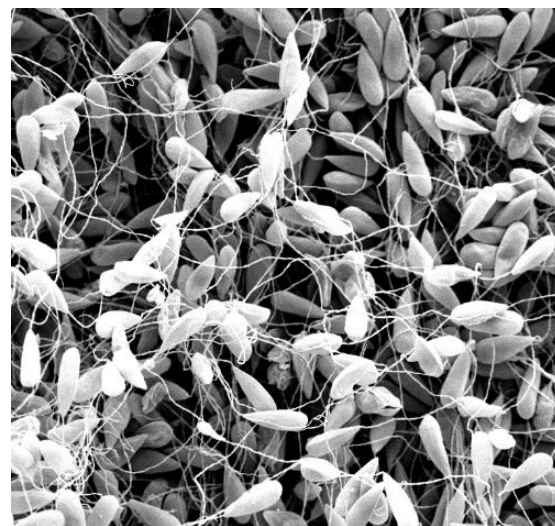
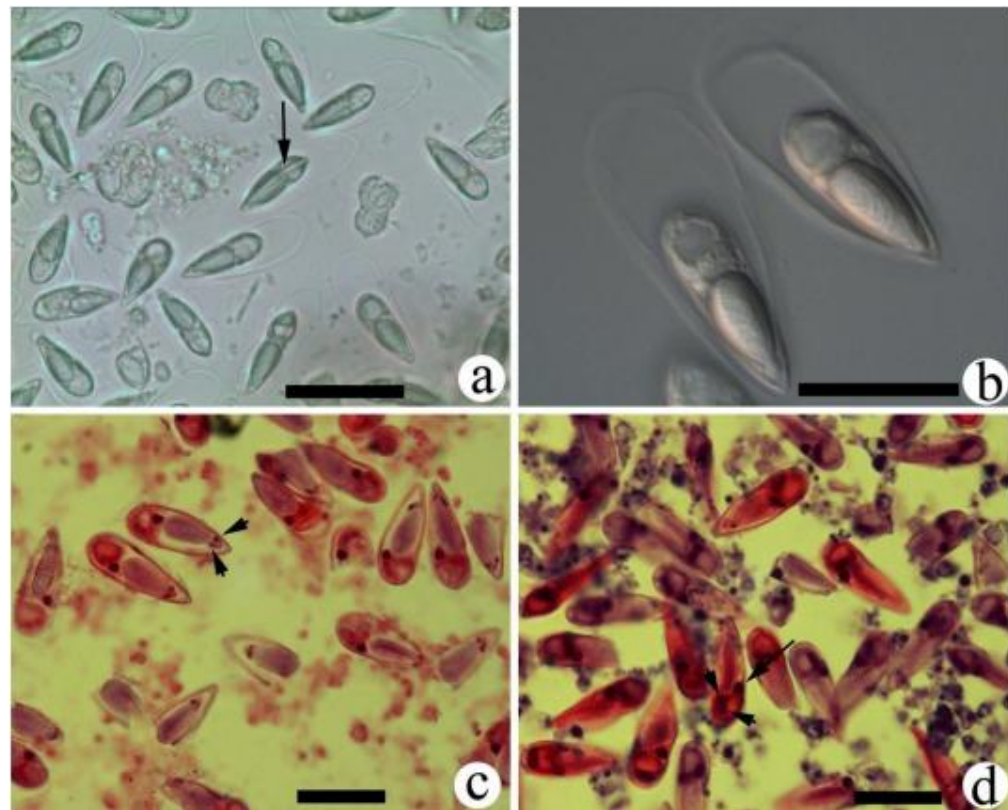
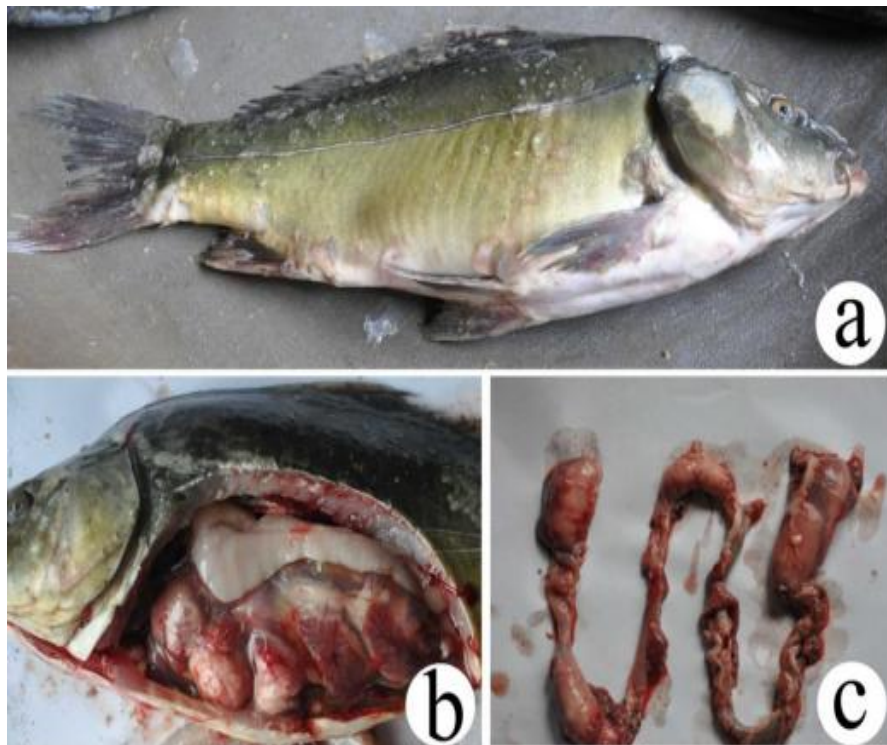


2.2 million tons/year;  
Fifth;  
100 species

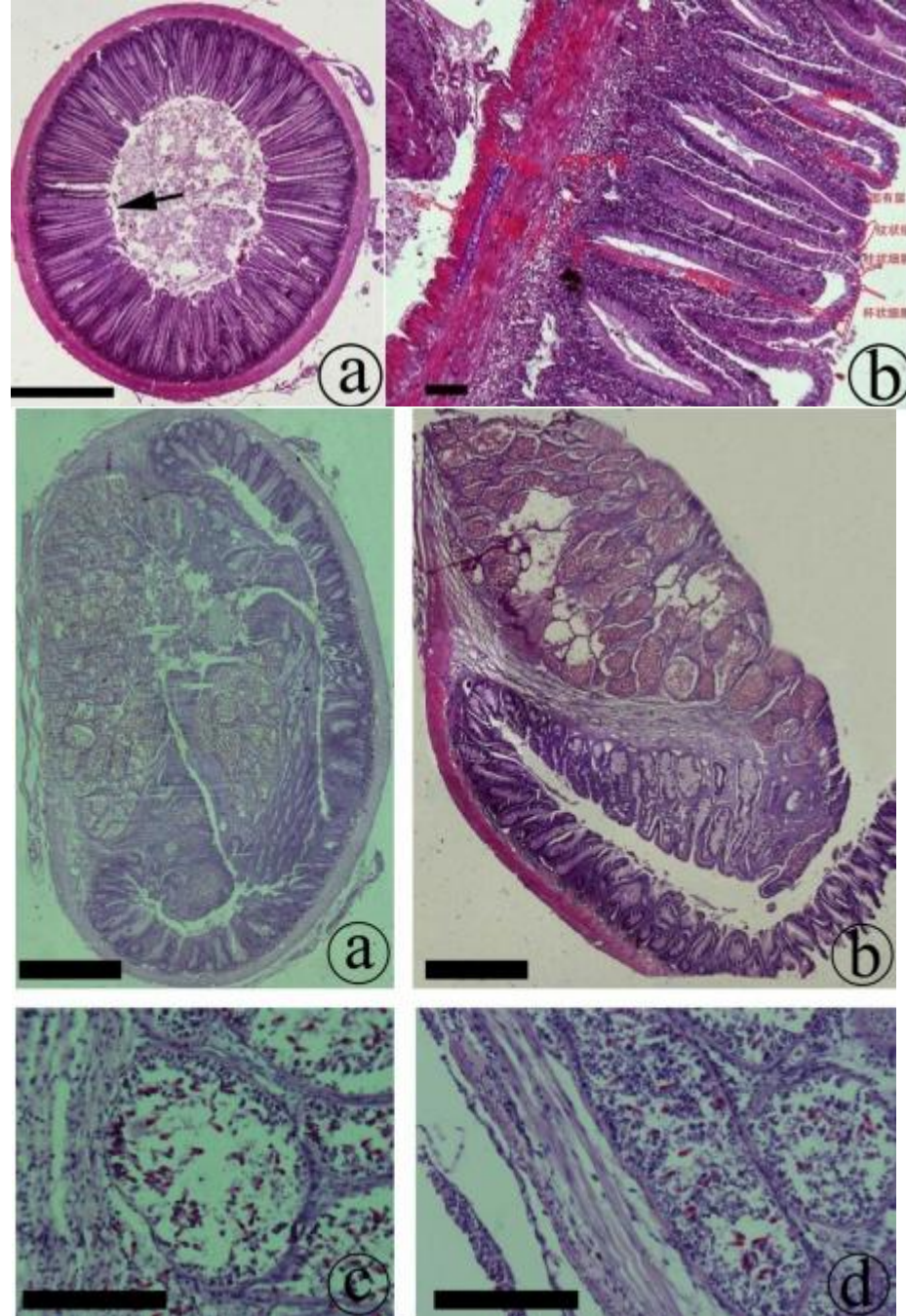
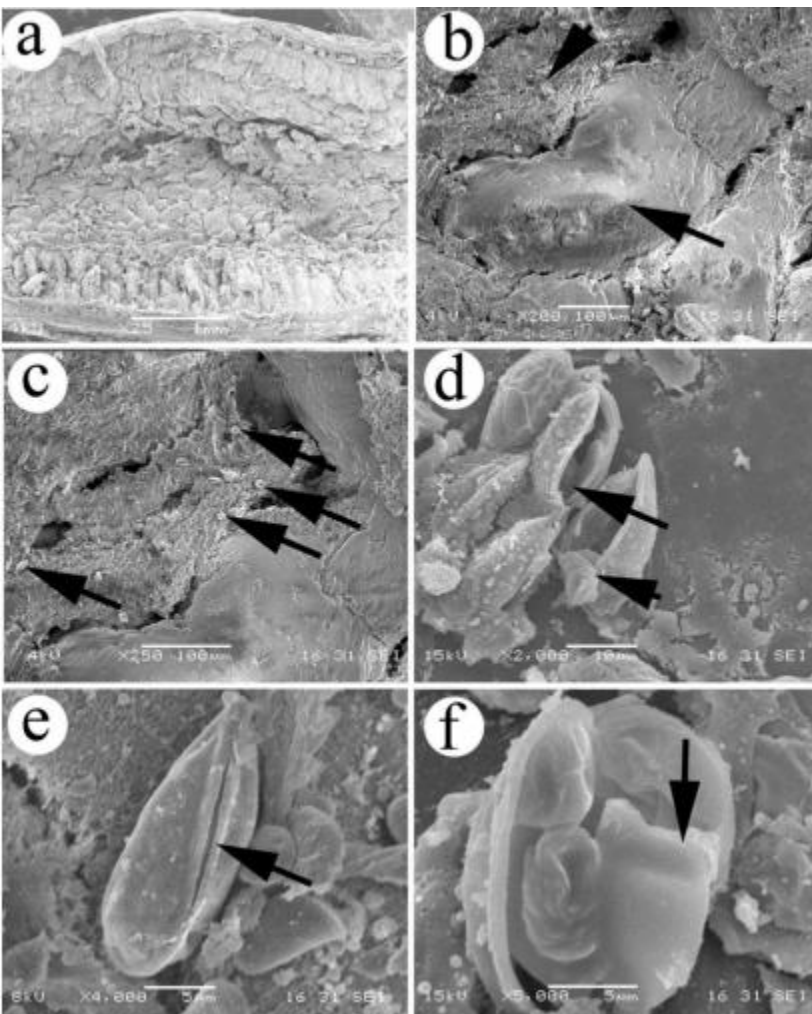
## Outline

- 1. Myxosporean infecting common carp**
- 2. Myxosporean infecting gibel carp**
- 3. Life cycles of fish myxosporean in China  
since 2009;**

*Thelohanellus kitauei*



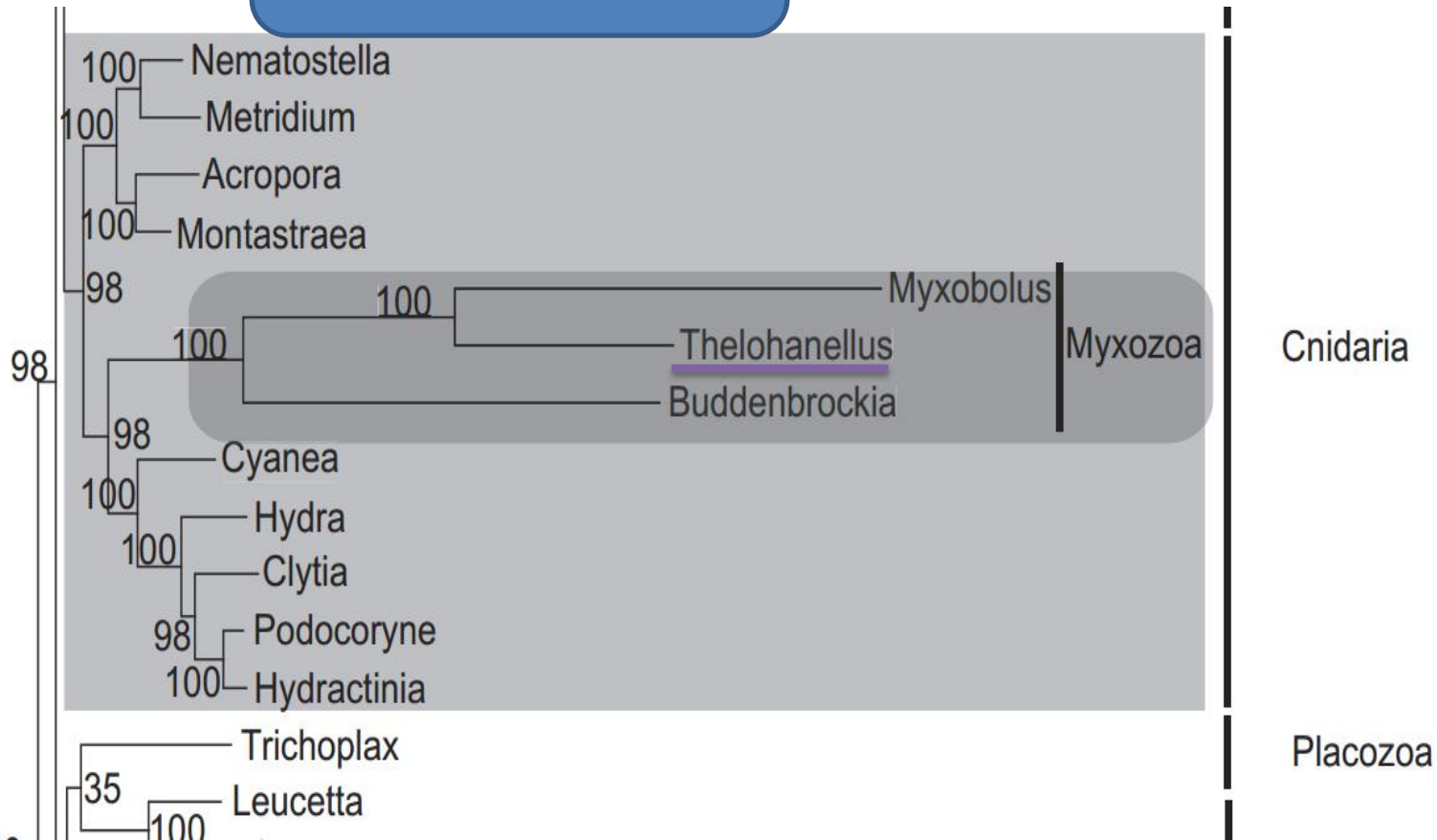
Clinical signs,  
Giant cystic diseases,  
also in Japan, Korea,  
Israel



# The Genome of the Myxosporean *Thelohanellus kitauei* Shows Adaptations to Nutrient Acquisition within Its Fish Host

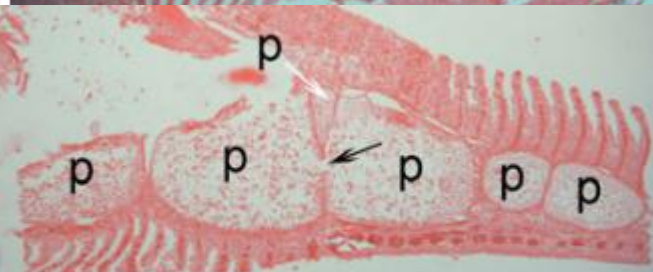
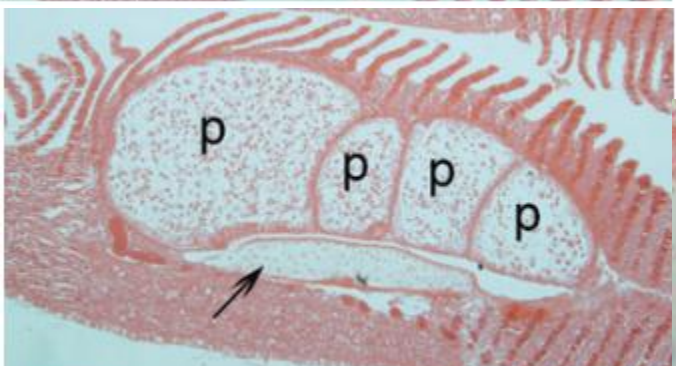
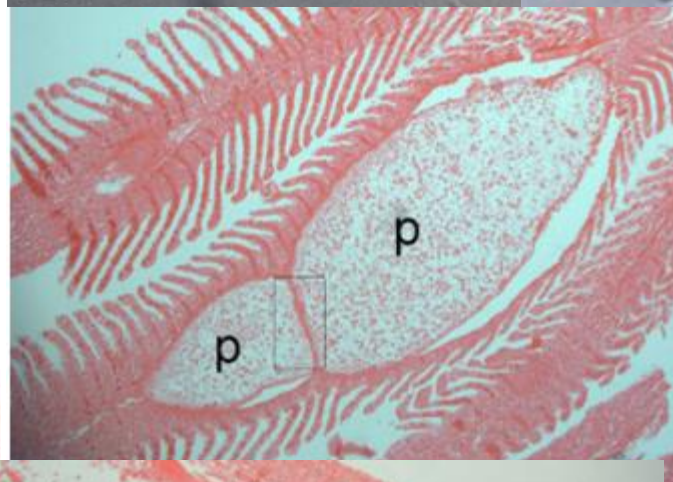
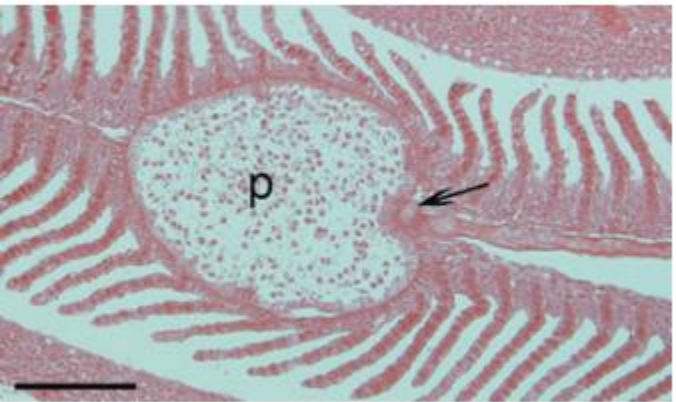
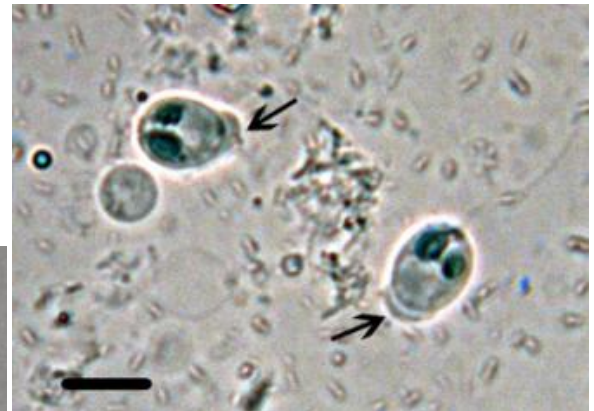
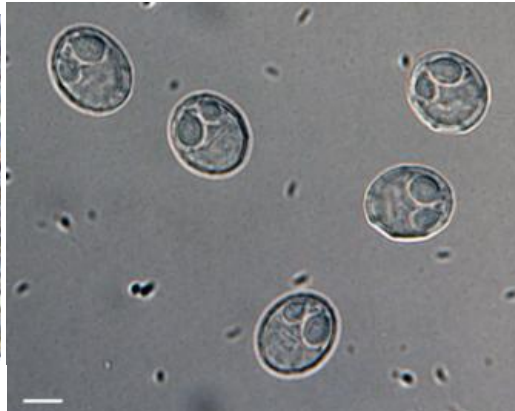
Yalin Yang<sup>1,†</sup>, Jie Xiong<sup>2,†</sup>, Zhigang Zhou<sup>1,\*</sup>, Fengmin Huo<sup>1</sup>, Wei Miao<sup>2</sup>, Chao Ran<sup>1</sup>, Yuchun Liu<sup>1</sup>,  
Jinyong Zhang<sup>2</sup>, Jin <sup>2</sup>, <sup>3</sup>, Wang<sup>4</sup>, Lei Wang<sup>4</sup>, and Bin Yao<sup>1,\*</sup>

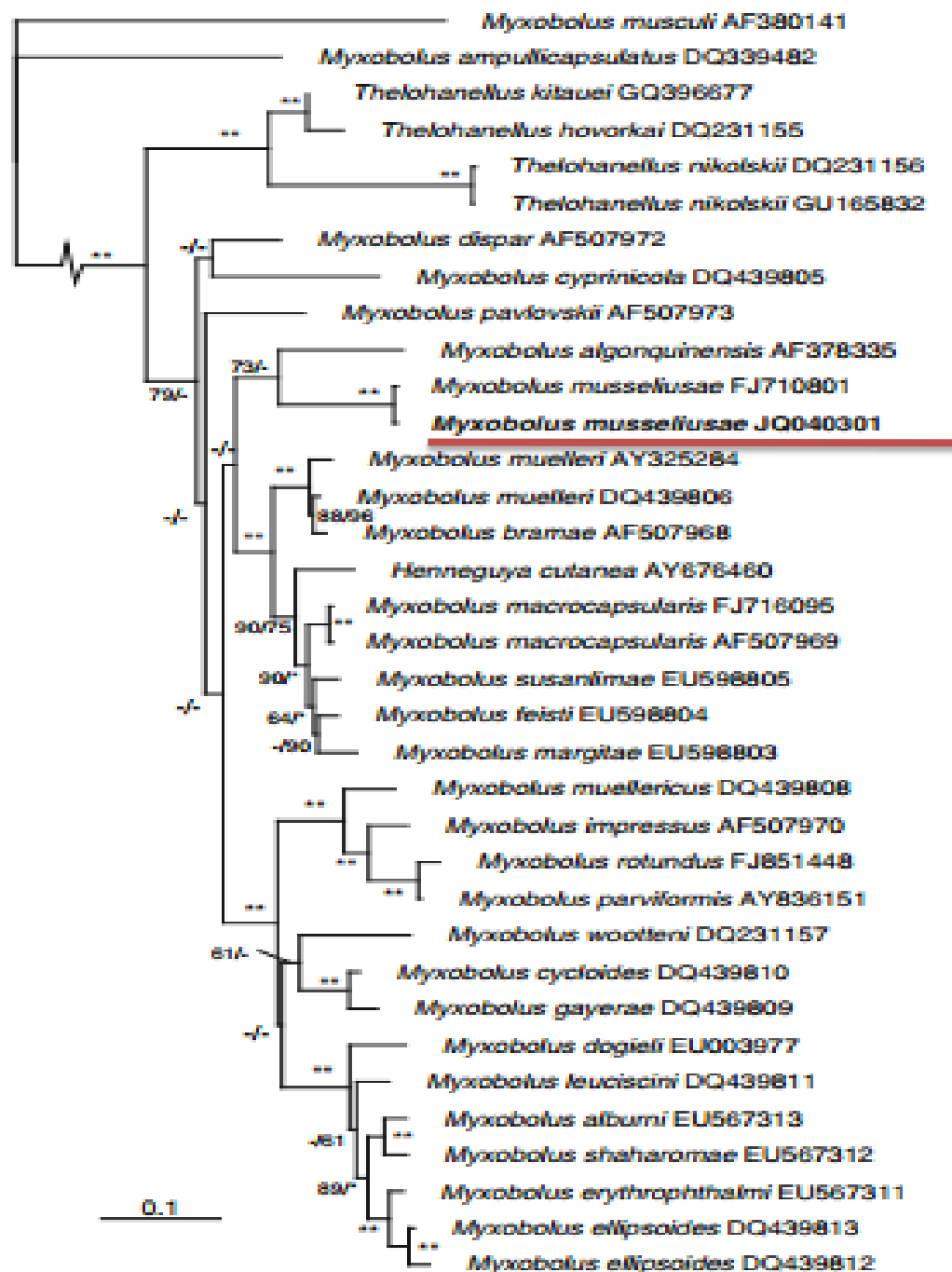
150.7 Mb, 5610 scaffolds



# *Myxobolus musseliusae*

Correction of *M. dispar* as *M. musseliusae* Yakovchuk, 1979  
Infection site: gill filament  
Molecular data: JQ040301



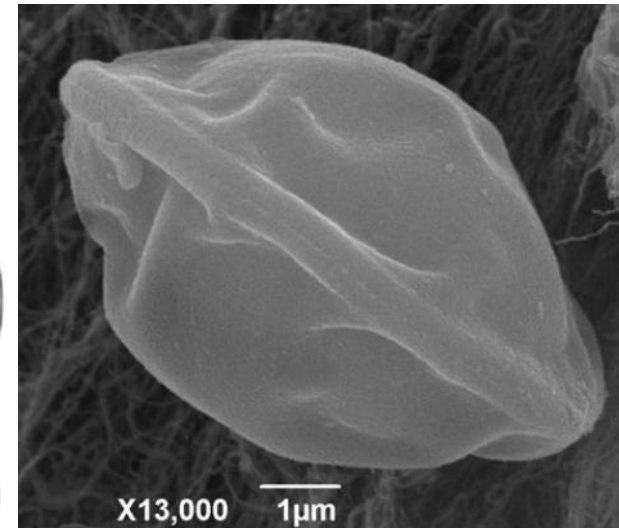
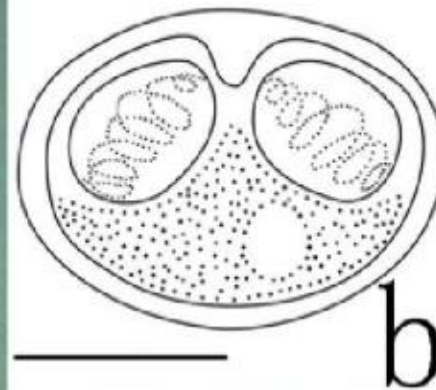
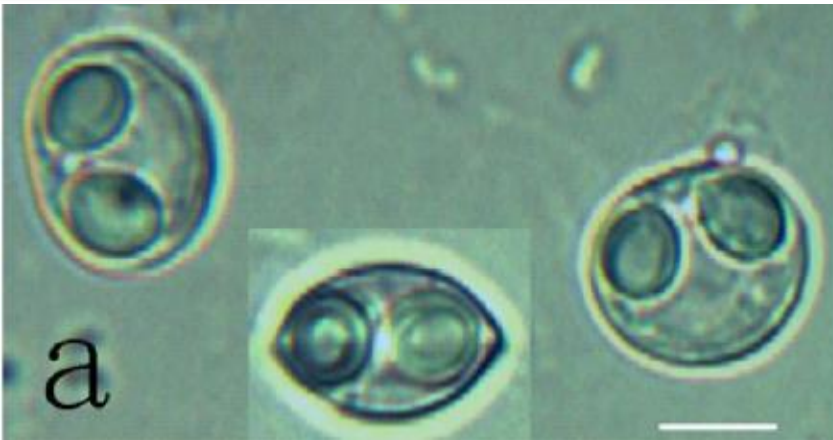


## *Myxobolus shangtungensis*

Redescription: *M. shantungensis* Hu, 1965;

Infection site: gill arche;

Molecular data: KJ725079

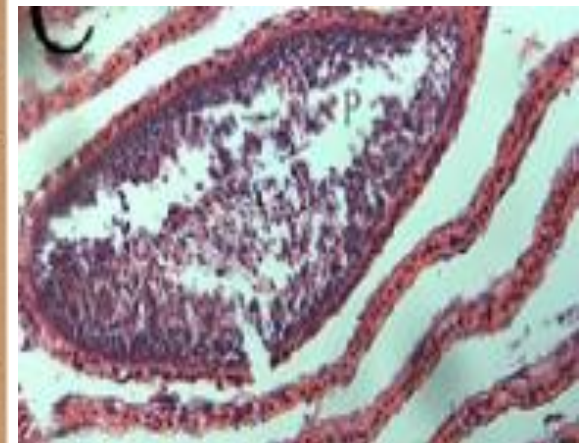
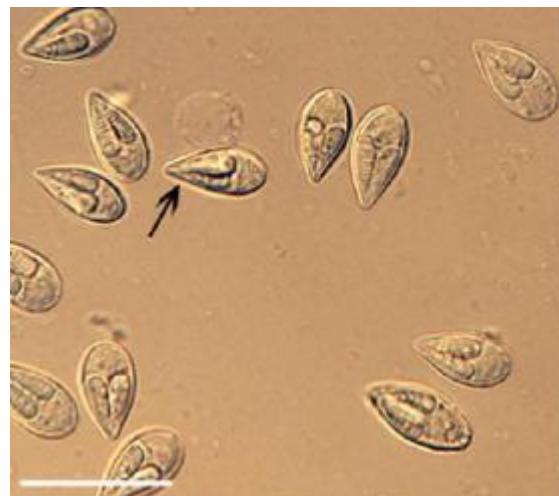


## *Myxobolus koi*

Redescription: *M. koi* Kudoa, 1919

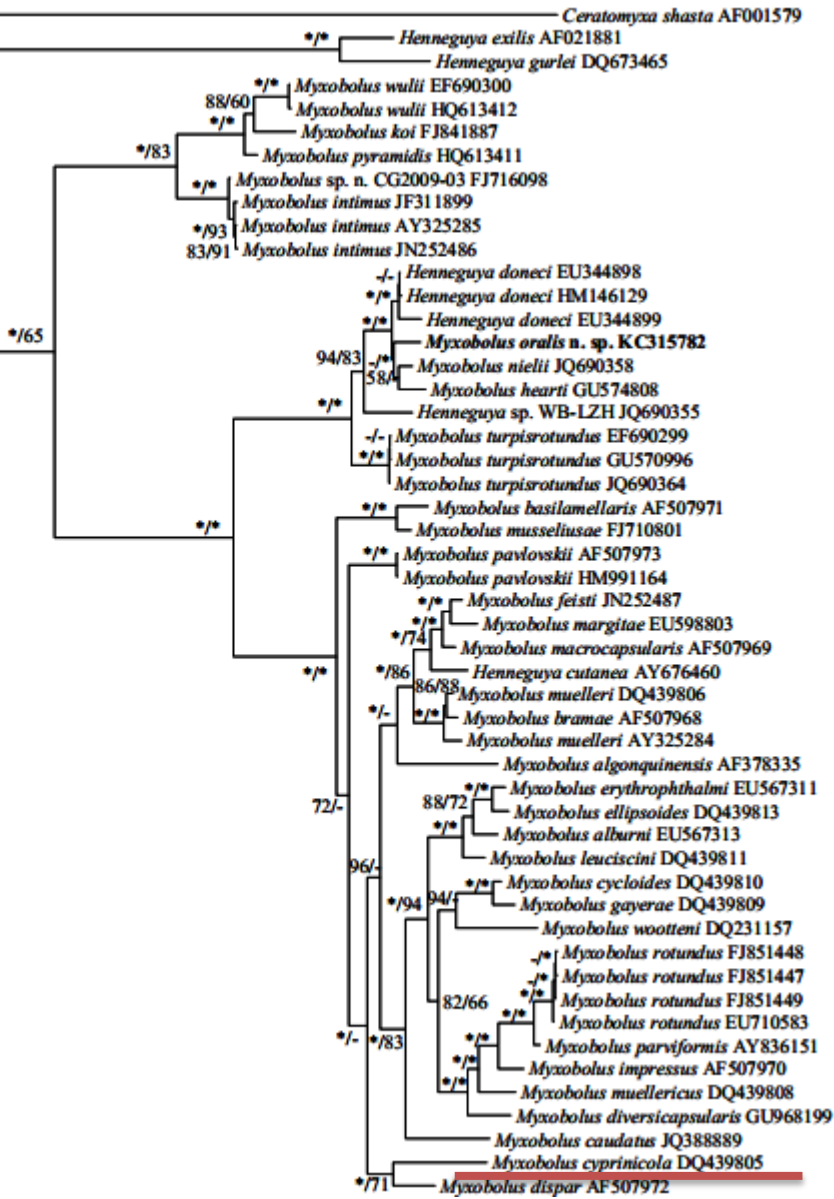
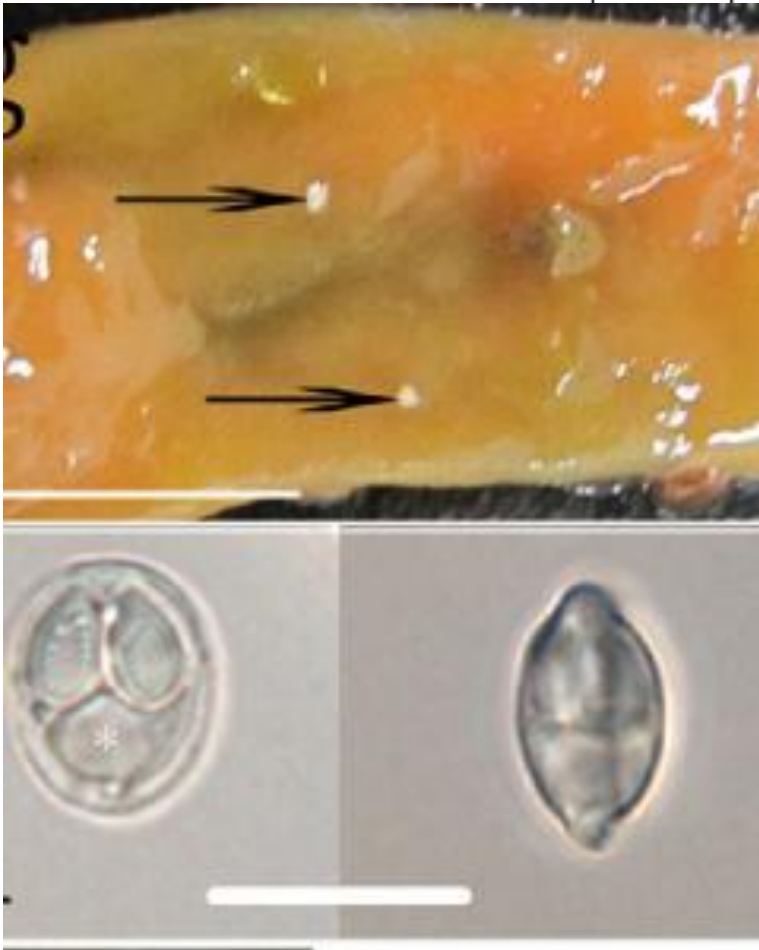
Infection site: gill lamellae;

Molecular data: KJ725077



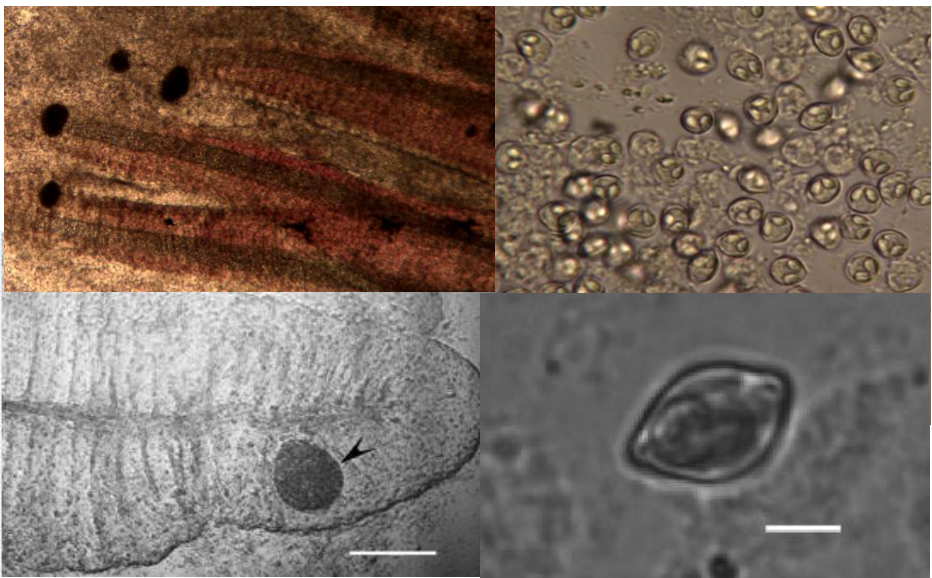
*Myxobolus cyprinicola*

Redescription: *M. cyprinicola* Reuss, 1966  
Infection site: intestine wall  
Molecular data: KJ725080

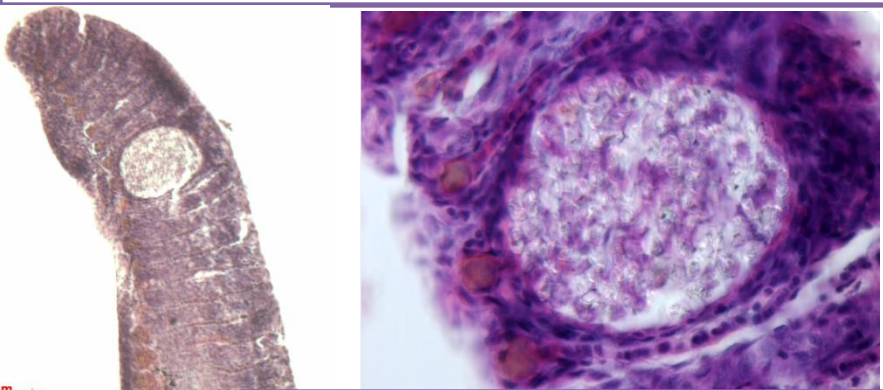


# Myxobolus pyramidis

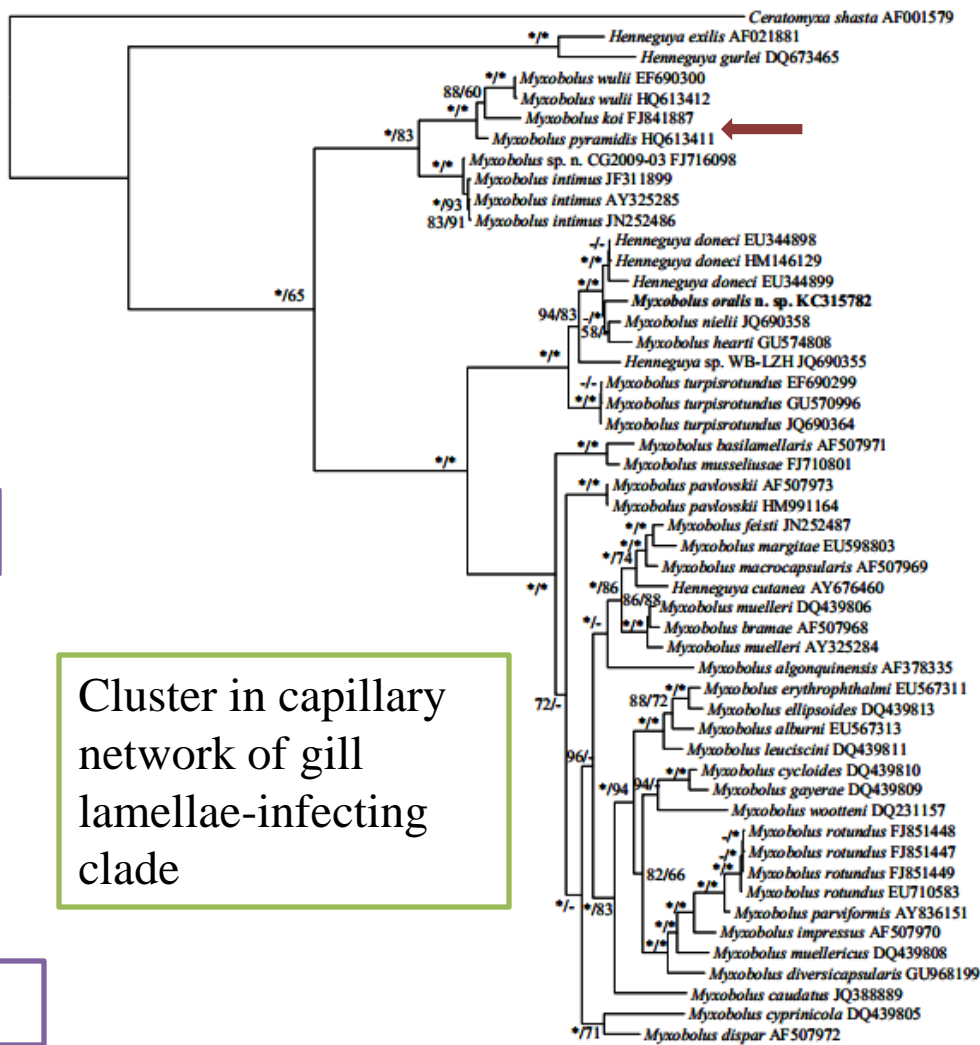
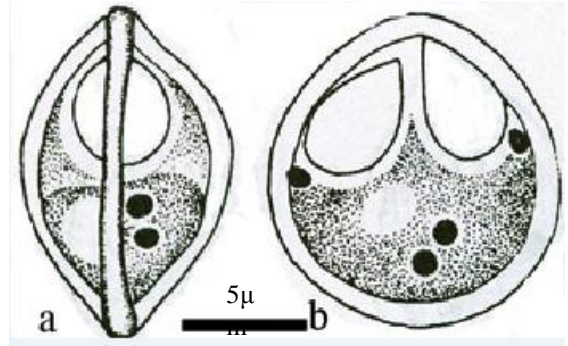
Redescription *M.pyramidis* Chen, 1958;  
Infection site: gill lamellae  
Molecular data: HQ613411



Showing plasmodium locating at the tip of gill filaments and spore in frontal and sutural view



Showing the location of plasmodium and no inflammatory responses besides mechanistic extrusion



Cluster in capillary network of gill lamellae-infecting clade

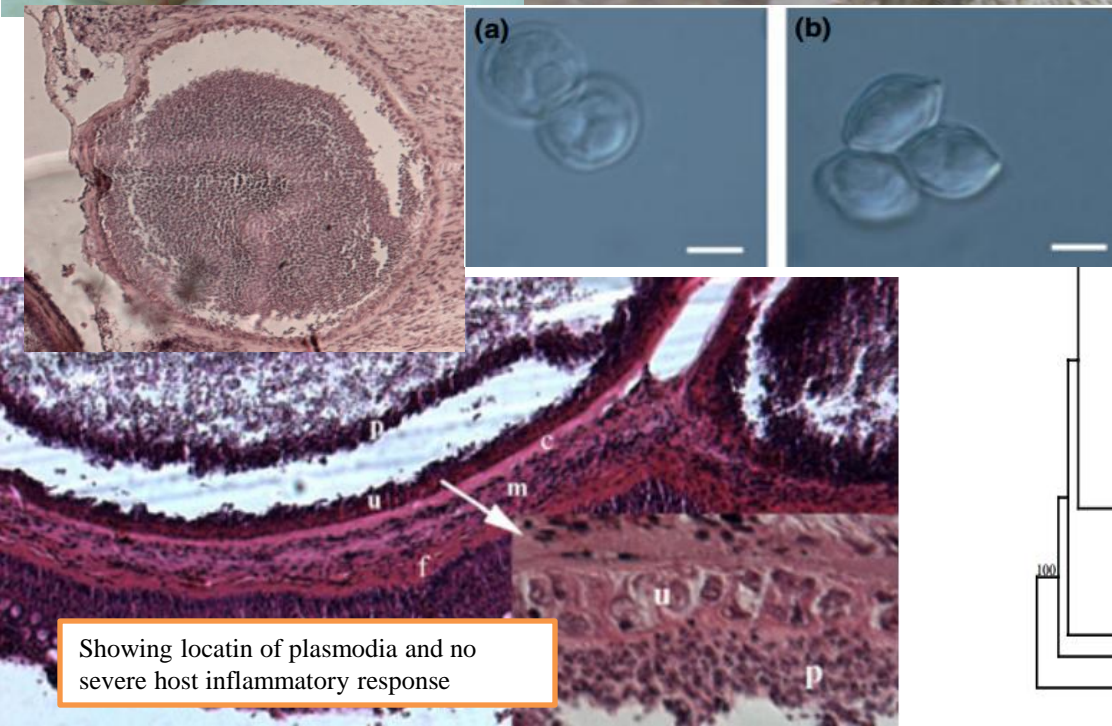
# *Myxobolus turpisrotundus* n.sp.

Correction of *M. rotundus* recorded in China;  
Infection site: connective tissue of gill arch, fins, intestine and body surface

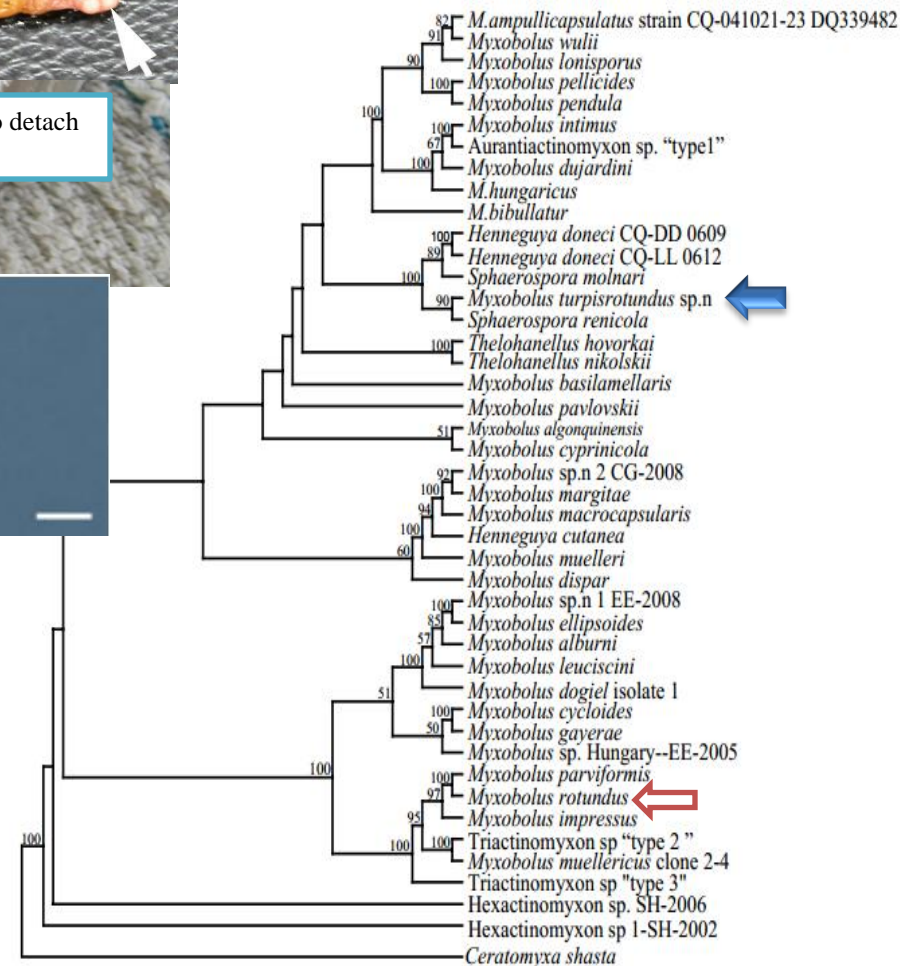
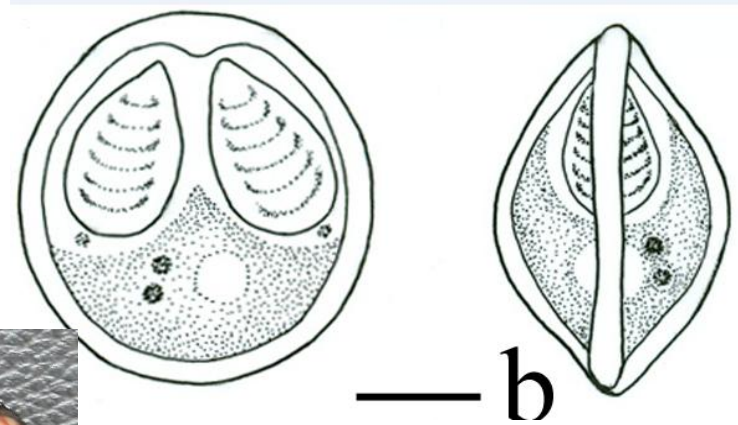
Molecular data: EF690299, GU570996



Plasmodium being going to detach from the dorsal fins



Showing locatin of plasmodia and no severe host inflammatory response

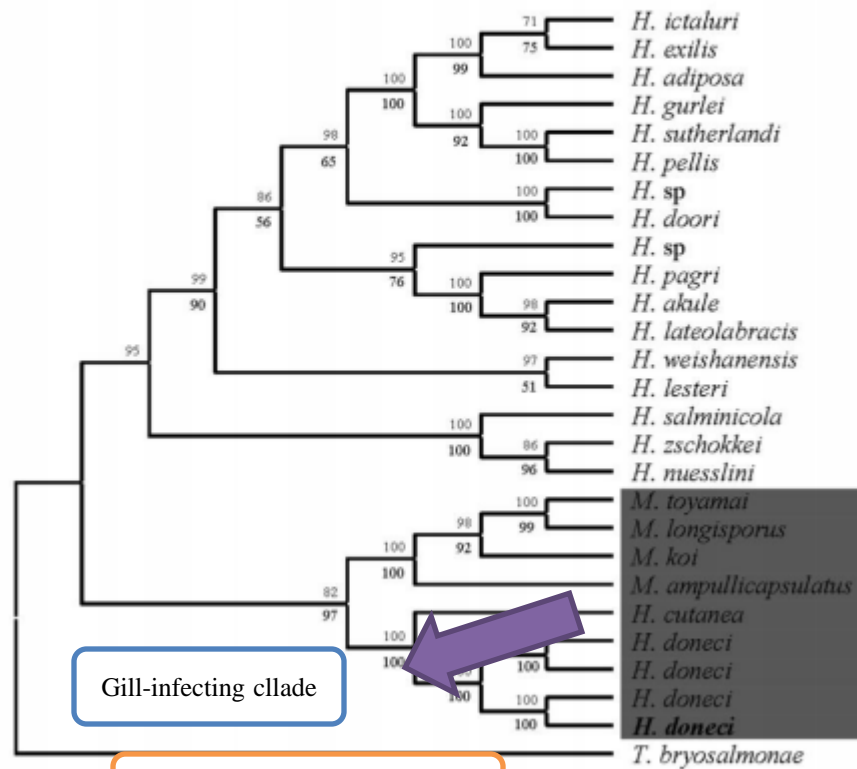
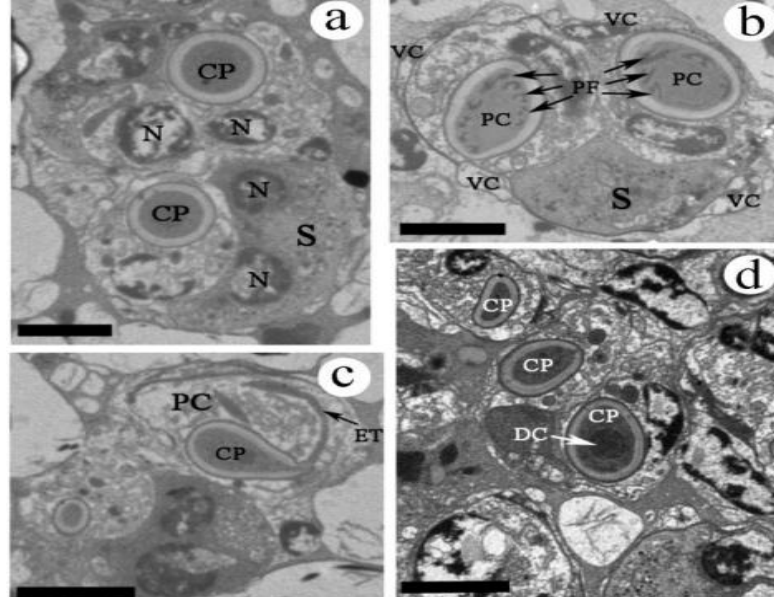
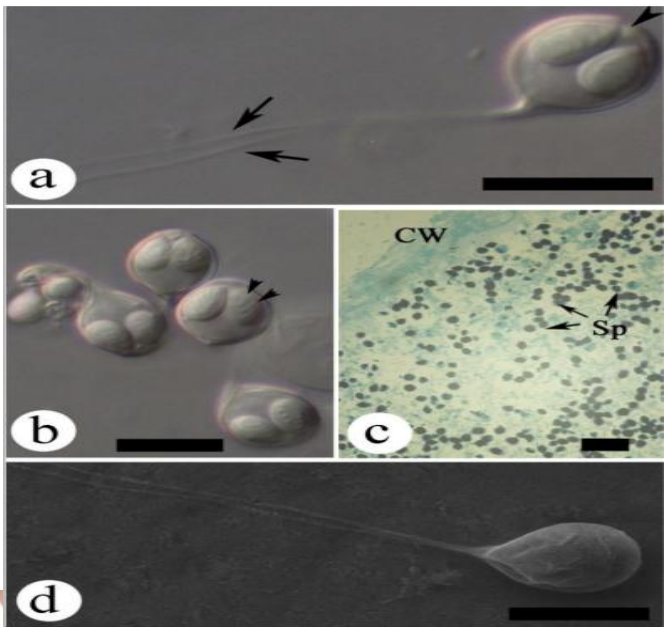


*Henneguya doneci* n. sp.

Redescription *M. doneci* Schulman, 1962;

Infection site: gill filaments

Molecular data: EU344898 ; HM146129; LC011456, JQ690376

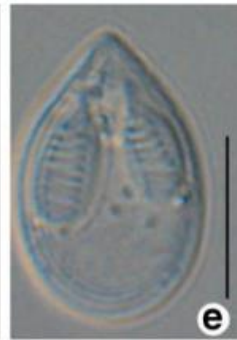
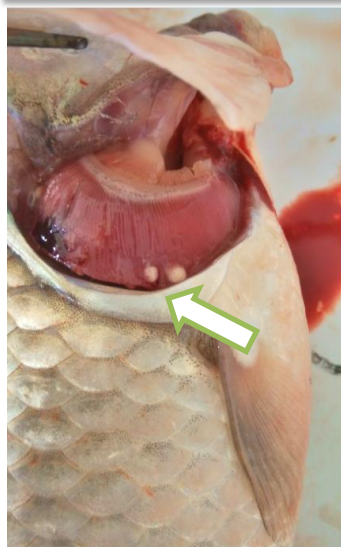


Cited from Ye et al. 2012

# *Myxobolus ampullicapsulatus* n. sp.

Infection site: gill filaments

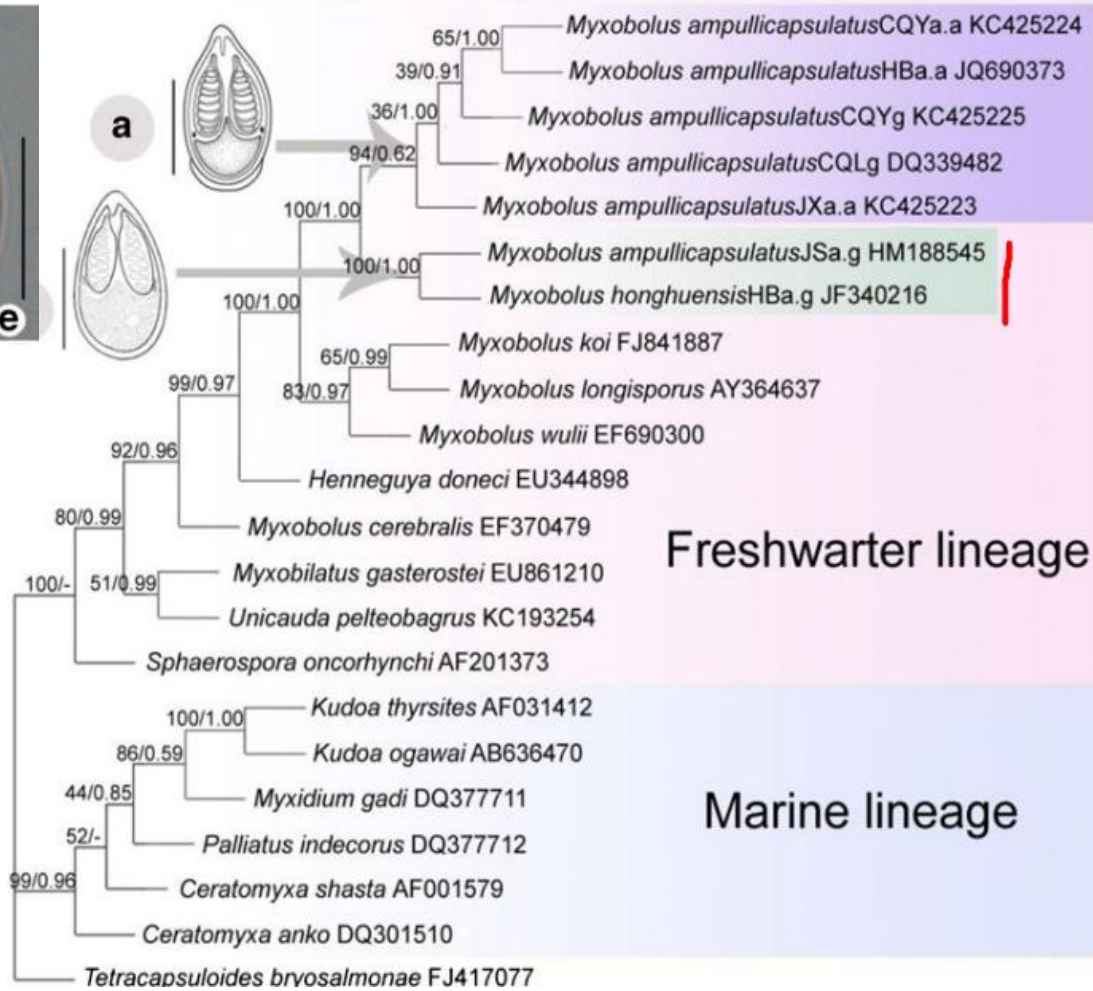
Molecular data: DQ339482, KJ725082, KC425225



*M. honghuensis*



*M. ampullicapsulatus*



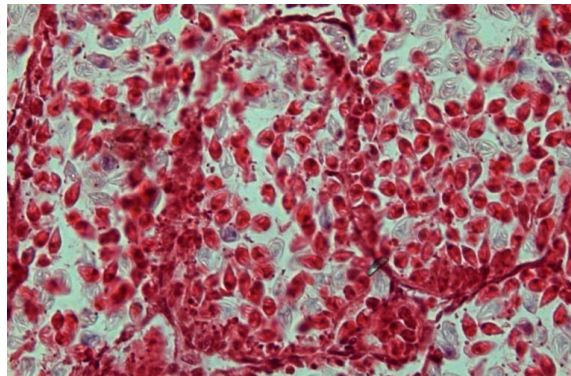
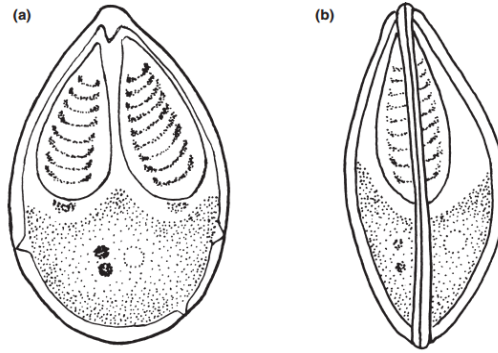
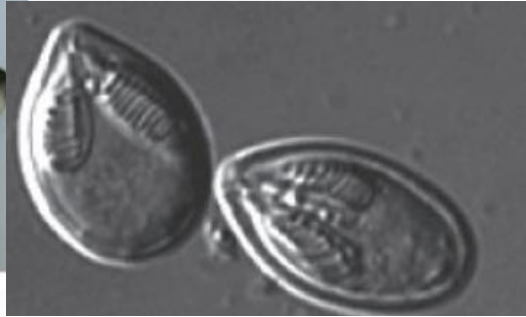
Showing tiny morphological and molecular difference between *M. ampullicapsulatus* and *M. honghuensis*

# *Myxobolus wulii*

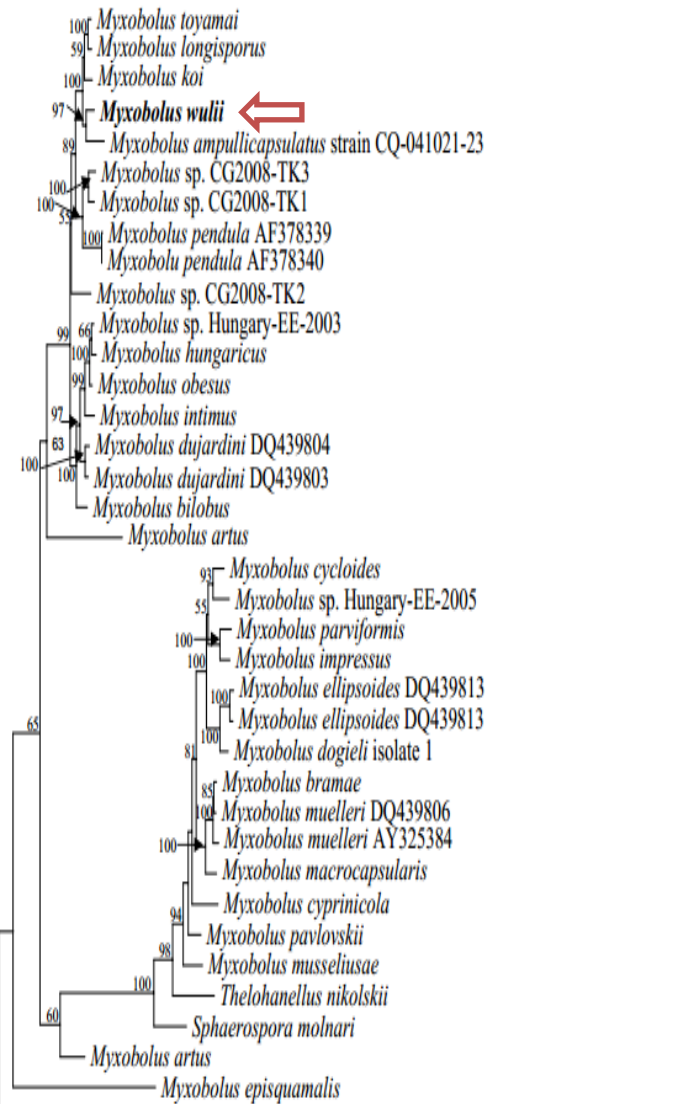
Synonyms: *Myxosoma magna* Wu & Li, 1986; *Myxobolus quanqiaoensis* Wu & Wang, 1997;

Infection site: hepatopancreas

Molecular data: EF690300



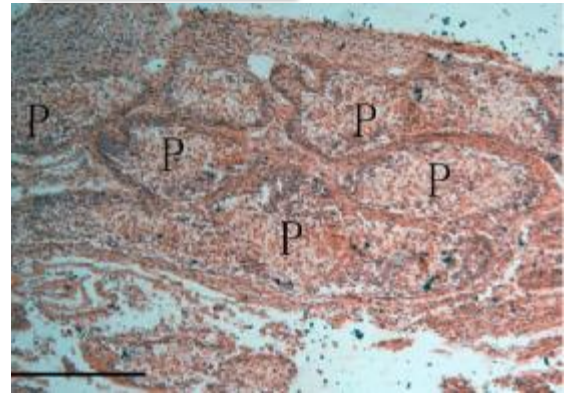
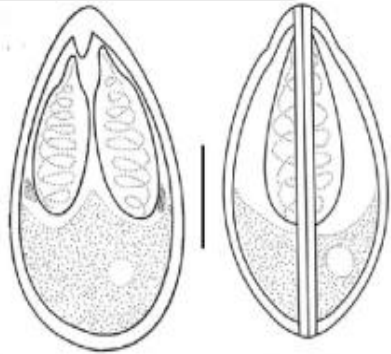
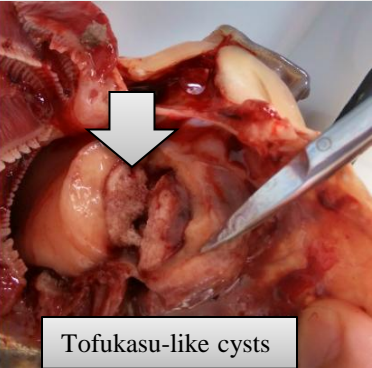
Honeycomb-like hepatocyte completely replaced by mature spores



*Ceratomyxa shasta* AF001579

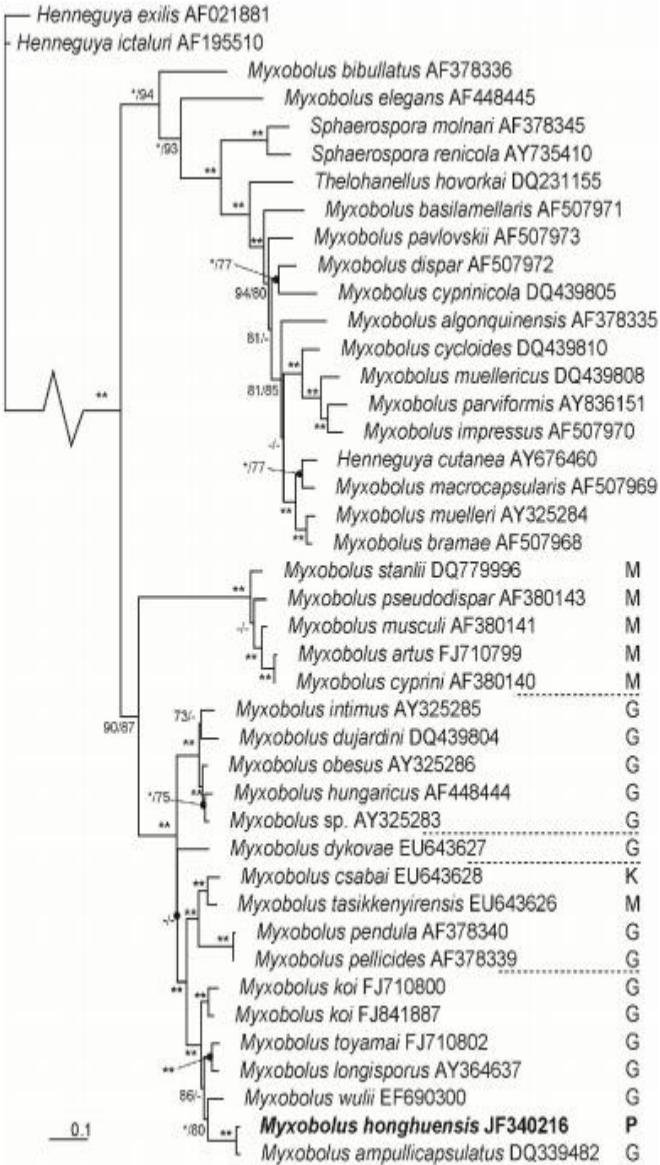
*Myxobolus honghuensis* n.sp.

Synonyms: *Myxosoma pharynx* Xiu & Lu 2013;*M. ampullicapsulatus* Jsa.a  
Infection site: pharynx  
Molecular data: KJ725074; JF340216



Many small plasmodia forming a big cyst

Parasite Source	<i>M. honghuensis</i> n. sp.	<i>M. wulii</i>	<i>M. ampullicapsulatus</i> Zhao et al. (2008)
Host	<i>C. auratus gibelio</i>	<i>C. auratus gibelio</i>	<i>C. auratus auratus</i>
Spore (body) shape	pyriform	pyriform	pyriform
Spore (body) length	16.9± 0.5 (15.1 - 17.8)	18.0± 0.8 (16.0 - 19.6)	18.0±0.9 (16.5-19.5)
Spore (body) width	10.4 ± 0.4 (9.8 - 11.3)	11.3 ± 0.6 (9.8 - 12.0)	9.3 ± 0.4 (8.5-10.0)
Spore (body) thickness	8.4 ± 0.4 (7.9 - 9.1)	8.9 ± 0.4 (7.8 - 9.8)	7.0
Large polar capsule length	8.4 ± 0.4 (7.6 - 9.2)	9.2 ± 0.6 (7.8 - 10.0)	
Large polar capsule width	3.9 ± 0.2 (3.1 - 4.2)	3.9 ± 0.2 (3.1 - 4.3)	8.5 ± 0.7 (7.0-10.0)
Small polar capsule length	7.9 ± 0.2 (7.0 - 8.3)	8.6 ± 0.5 (7.5 - 10.0)	
Small polar capsule length	3.7 ± 0.3 (3.0 - 4.1)	3.8 ± 0.3 (3.0 - 4.2)	3.0 ± 0.2 (2.5 - 4.0)
No. filament turns	7-8	7-9	9 - 10



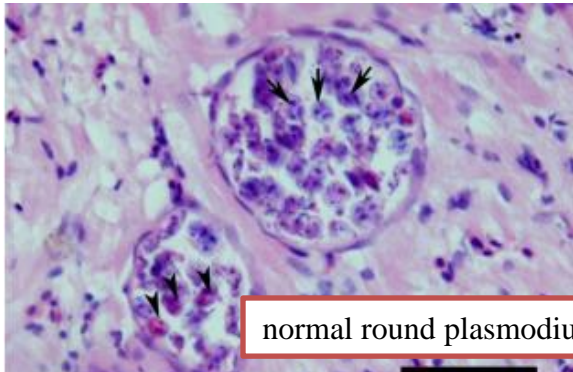
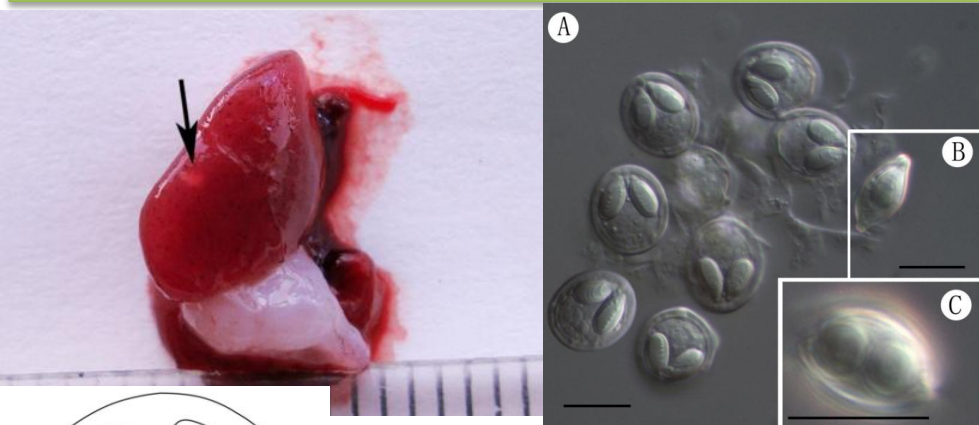
Cluster with species with elongate spore

# Myxobolus hearti

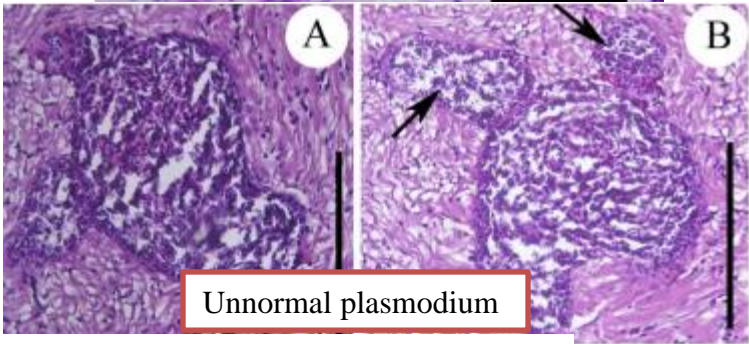
Redescription

Infection site: myocardium

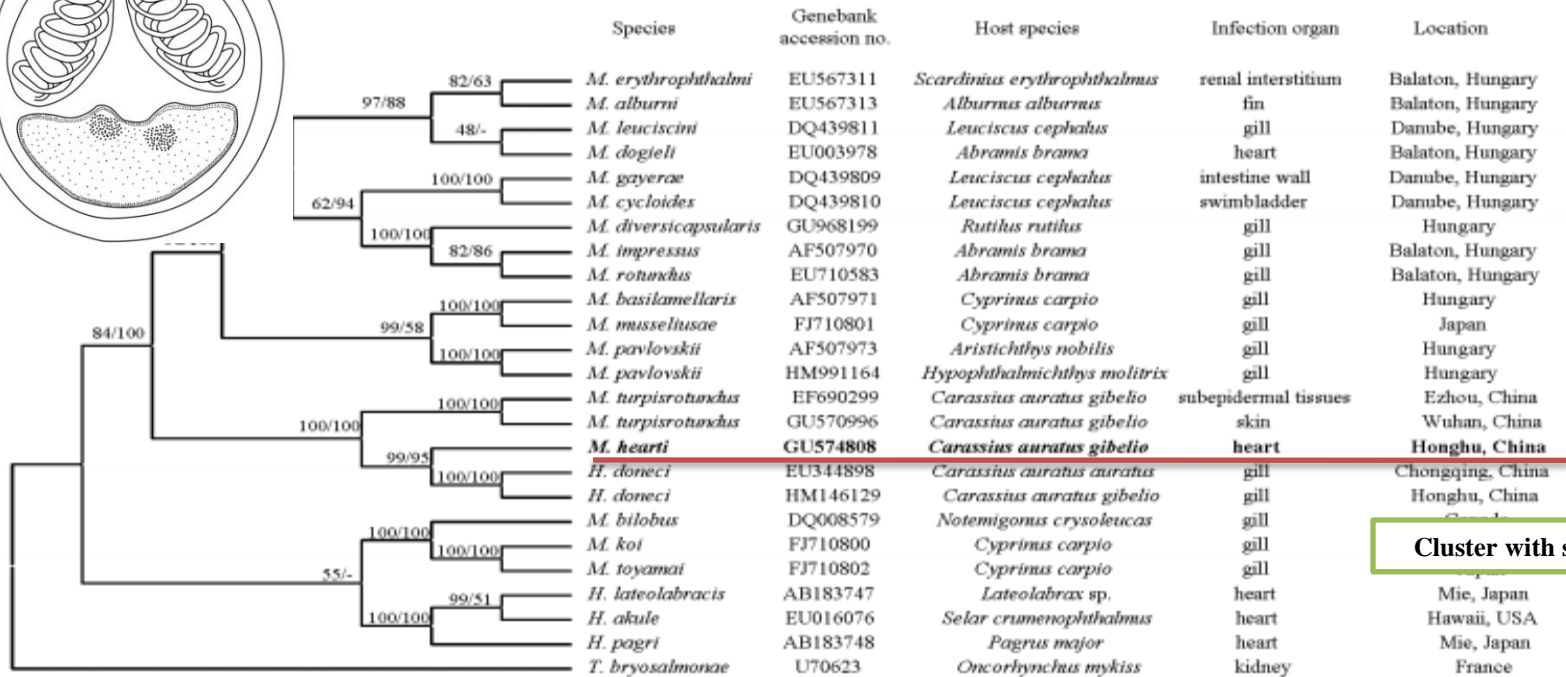
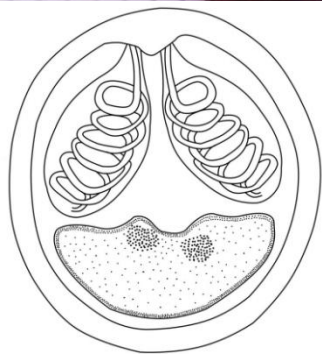
Molecular data: GU574808



normal round plasmodium



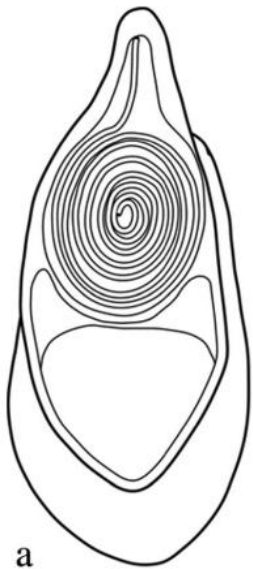
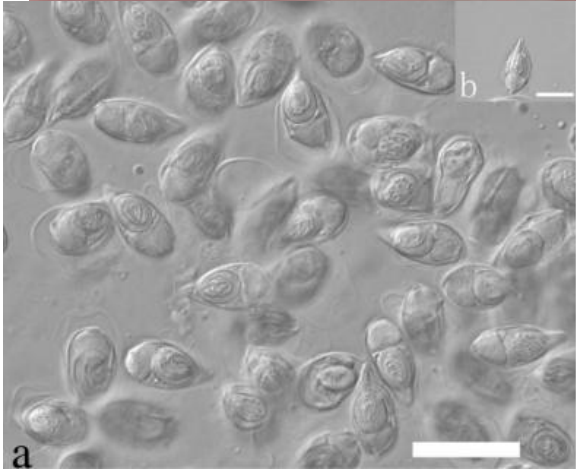
Unnormal plasmodium



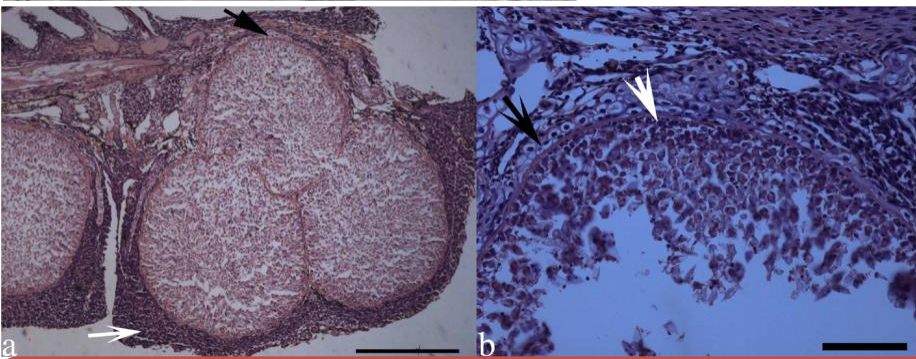
Cluster with species with round spore

*Thelohanellus wangi* n.sp.

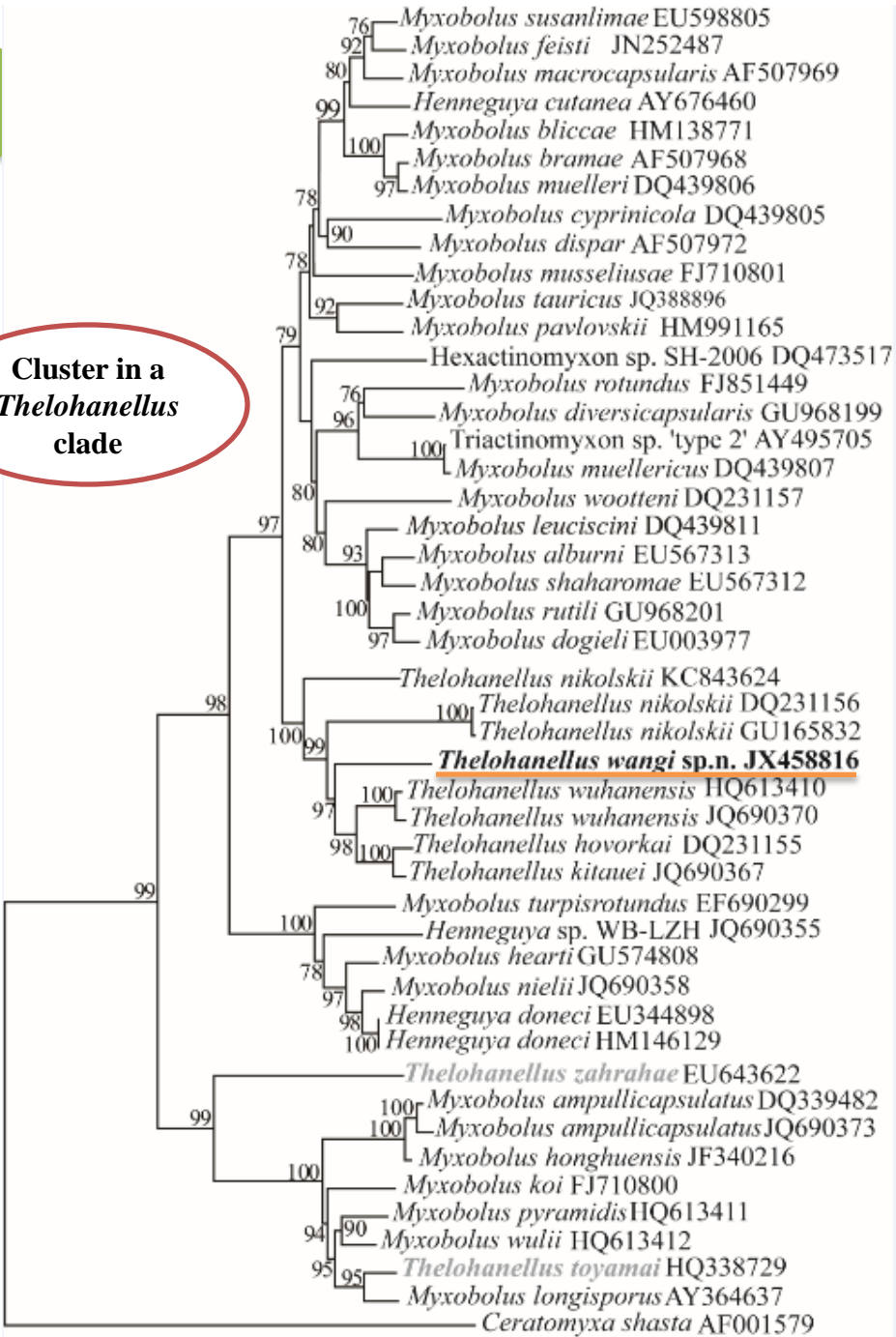
Infection site: gill filaments  
Molecular data: JX458816



Cluster in a  
*Thelohanellus*  
clade



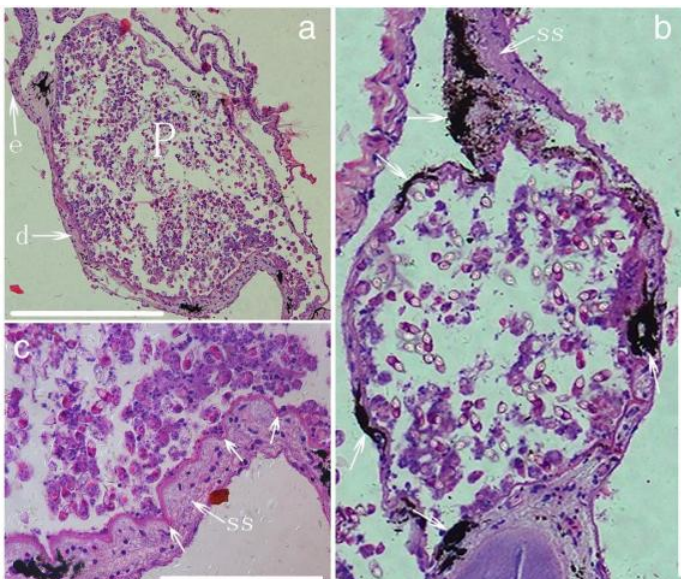
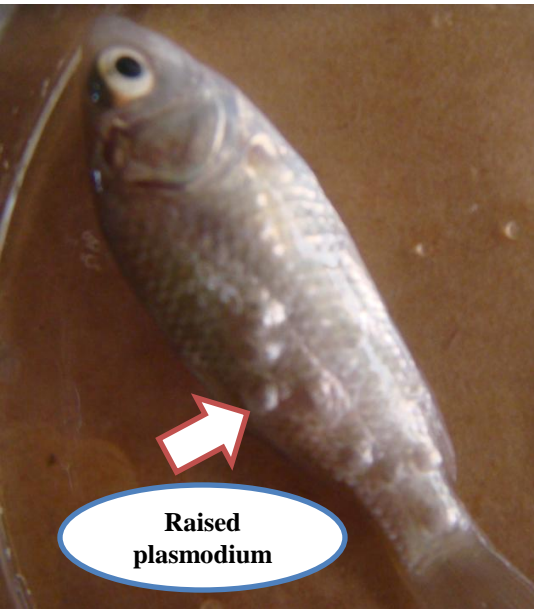
Obvious epithelial hyperphasia at the anterior part of infected gill filaments and a thin connective tissue membrane delimited the plasmodium from cartilage cells of gill filaments



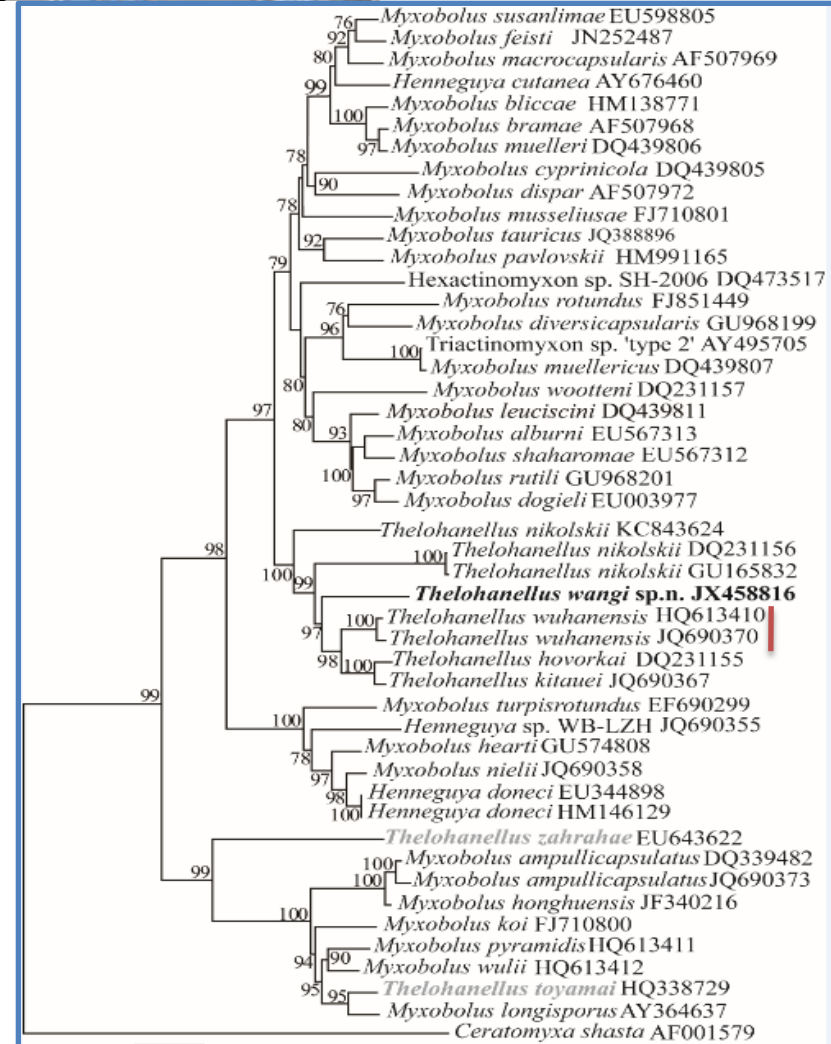
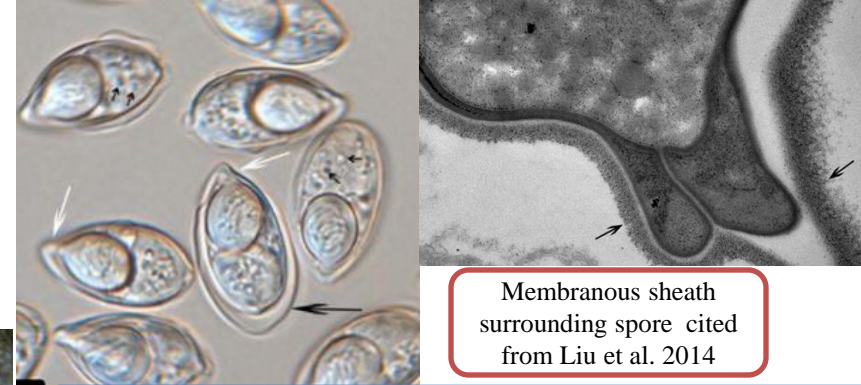
# *Thelohanellus wuhanensis* n. sp.

Infection site: skin

Molecular data: JQ968687; JQ088179; HQ613410

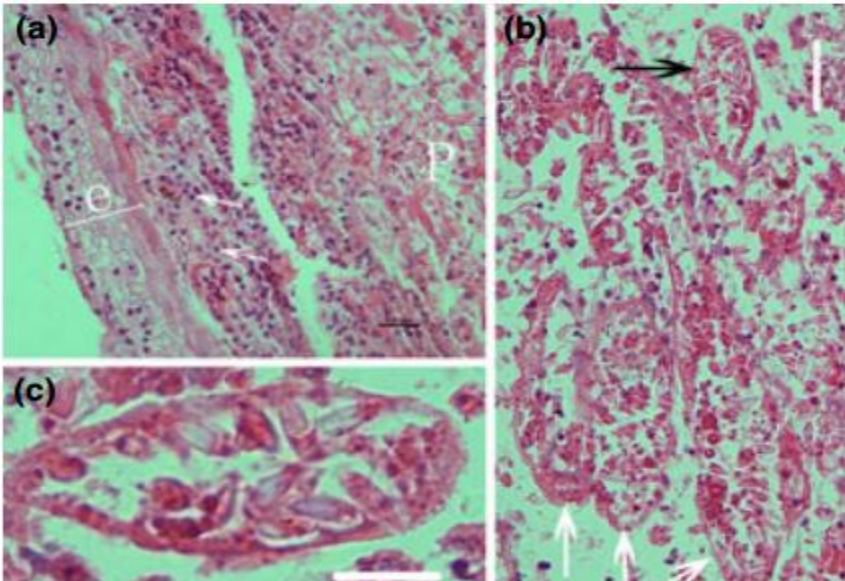
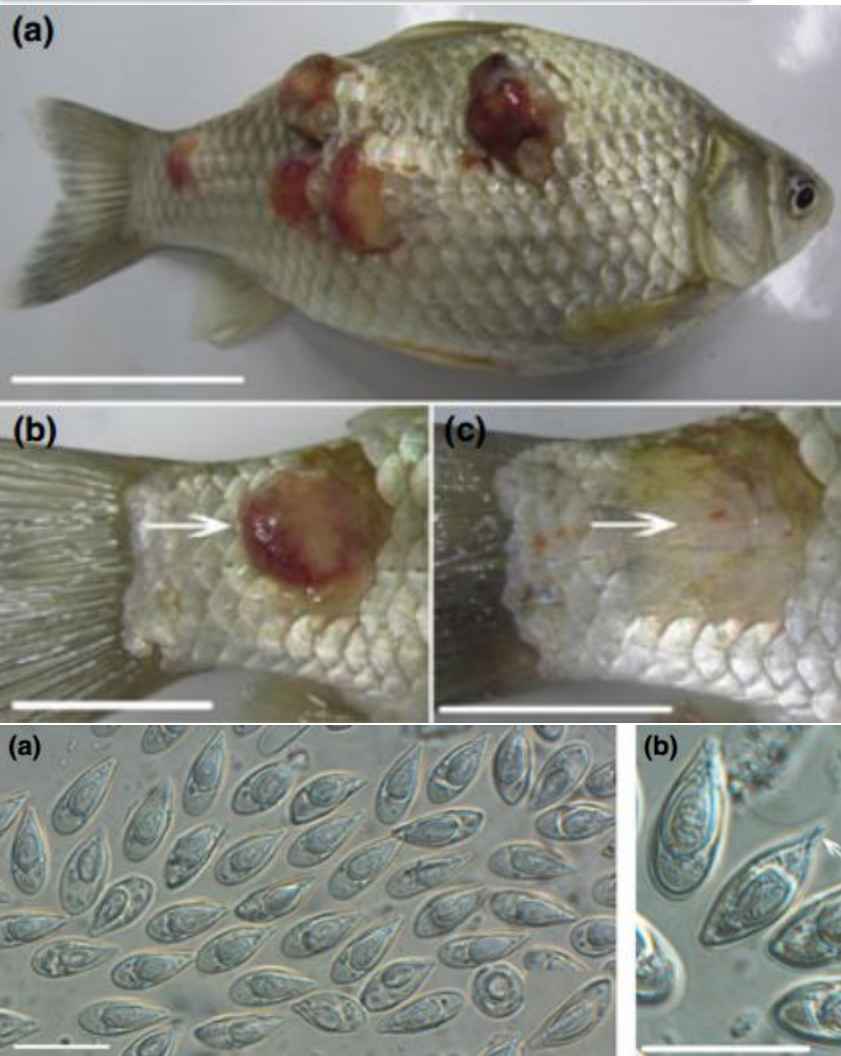


Showing plasmodium developing in the stratum spongiosum of dermis and many melanocytes gathering together around the plasmodia

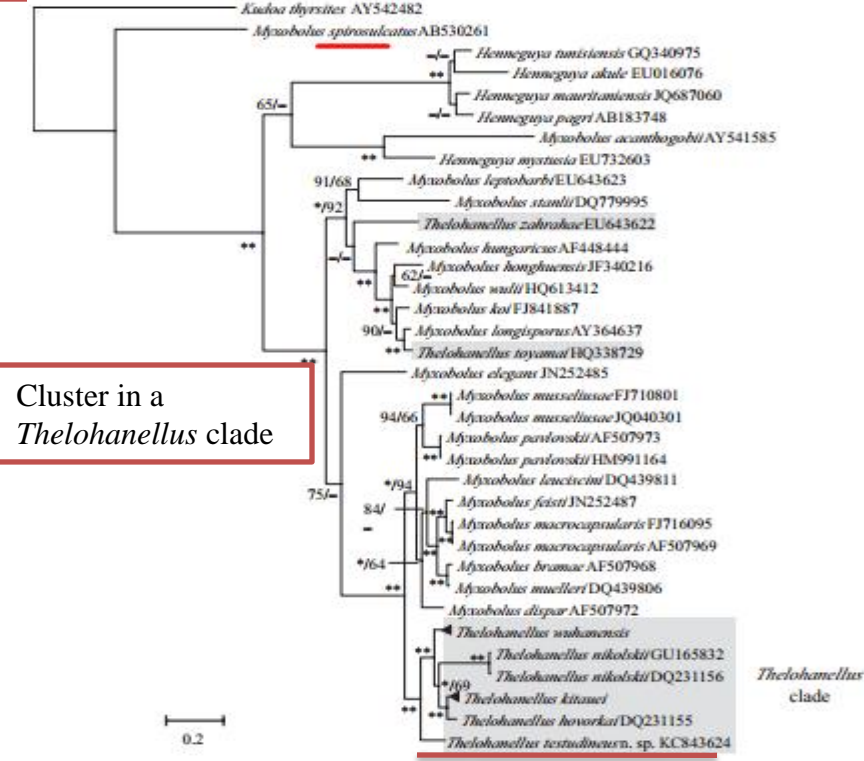


*Thelohanellus testudineus* n. sp.

Infection site: skin  
Molecular data: KC843624

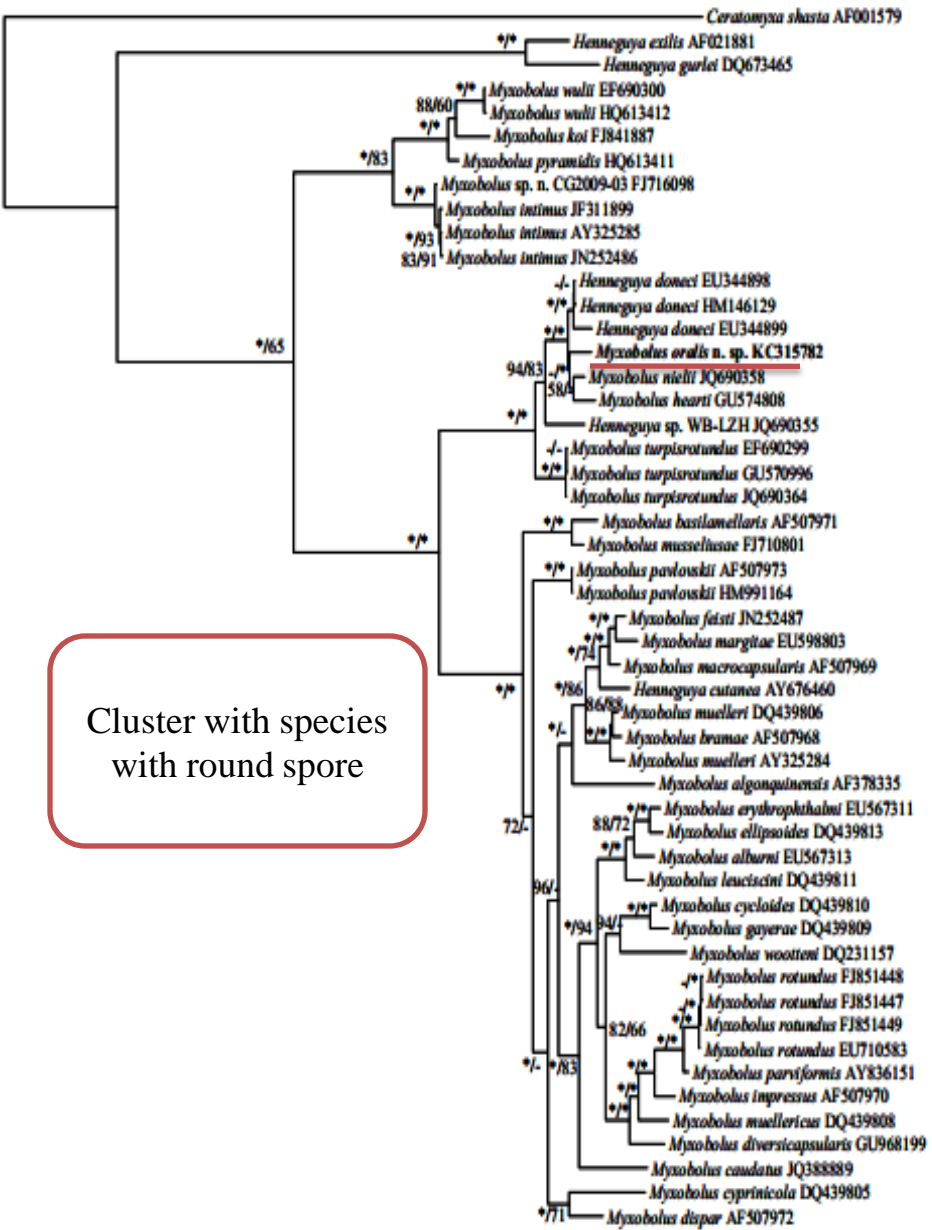
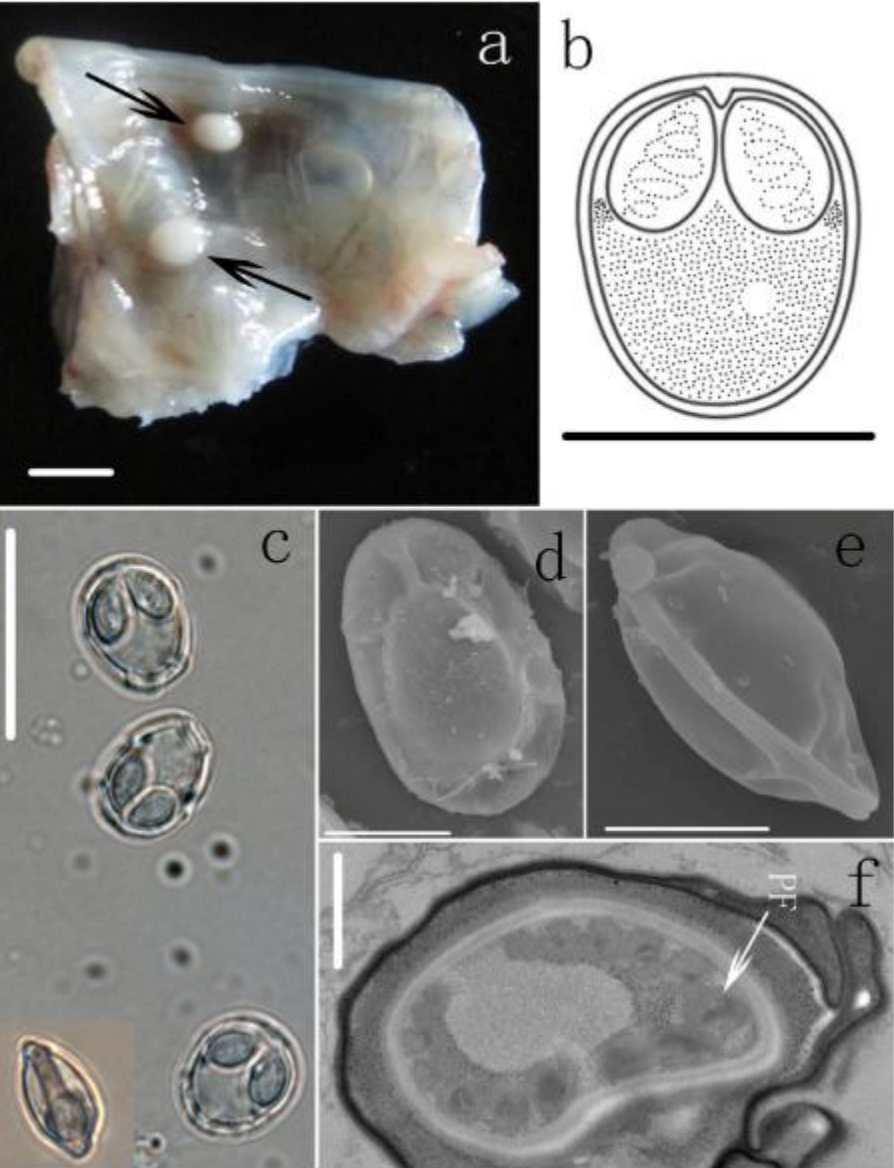


Showing plasmodium developing in the dermis of the skin and distinct inflammatory infiltration,



*Myxobolus oralis* n. sp.

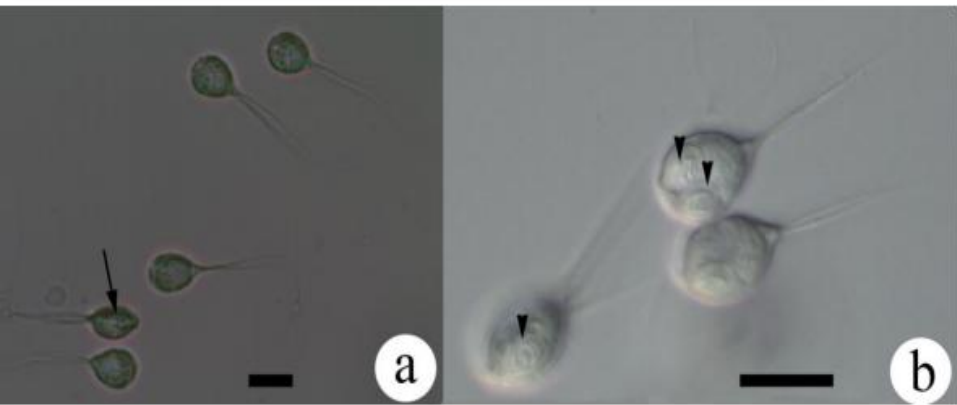
Infection site: oral cavity  
Molecular data: KC315782



Cluster with species  
with round spore

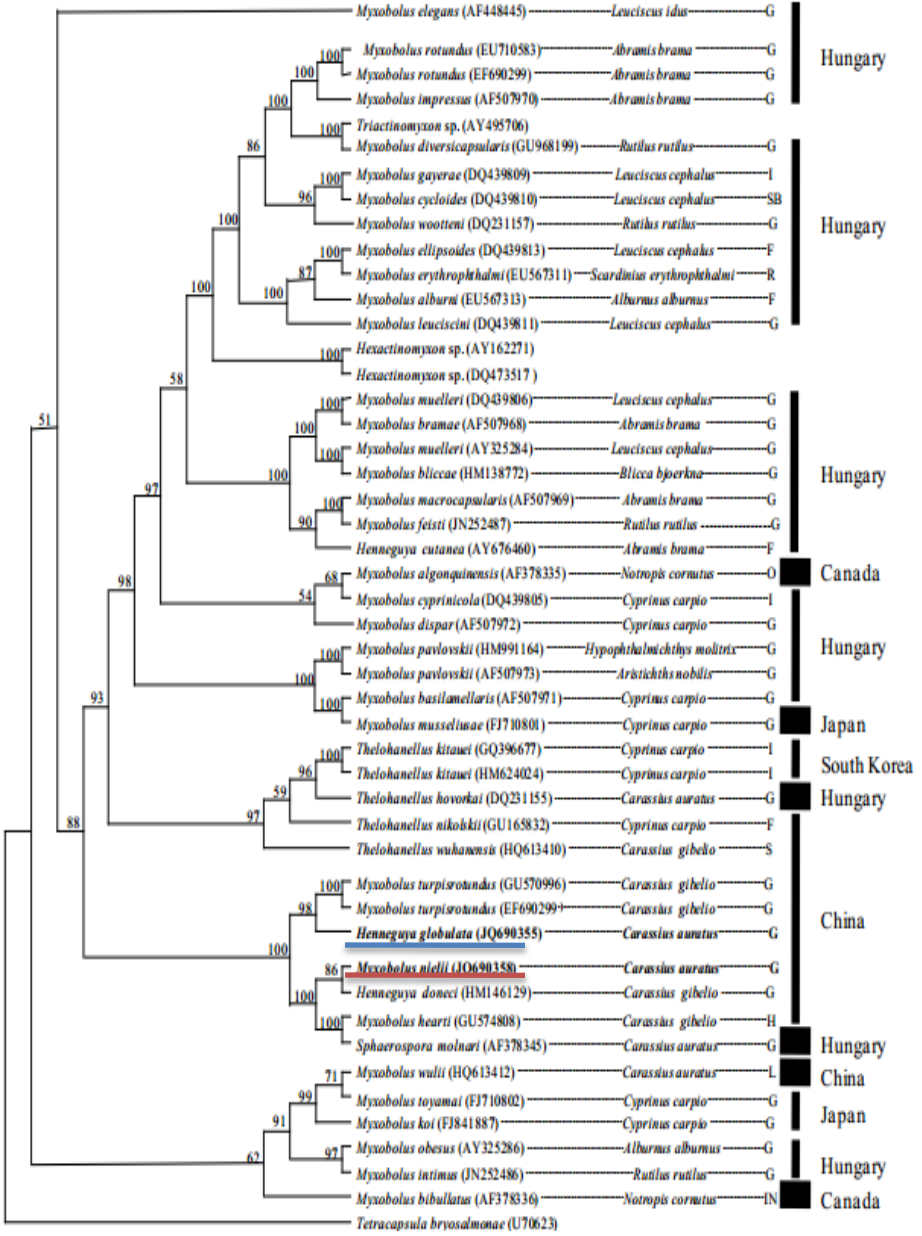
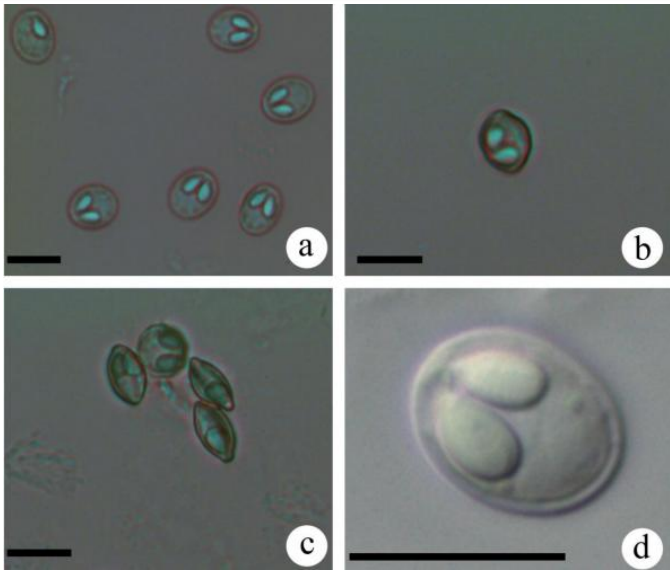
*Henneguya globulata* n.sp.

Infection site: gill filaments  
Molecular data: KJ725074; JF340216



*Myxobolus neli* (Nie et Li, 1973) Landsberg et Lom, 1991

Infection site: gill filaments  
Molecular data: KJ725074; JF340216

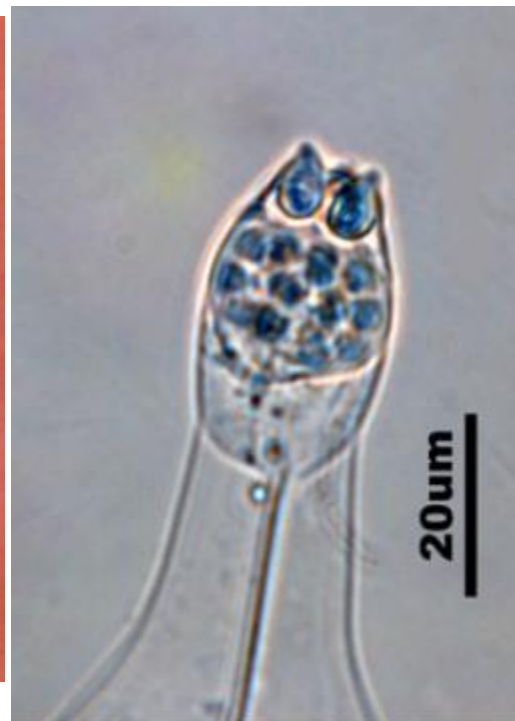
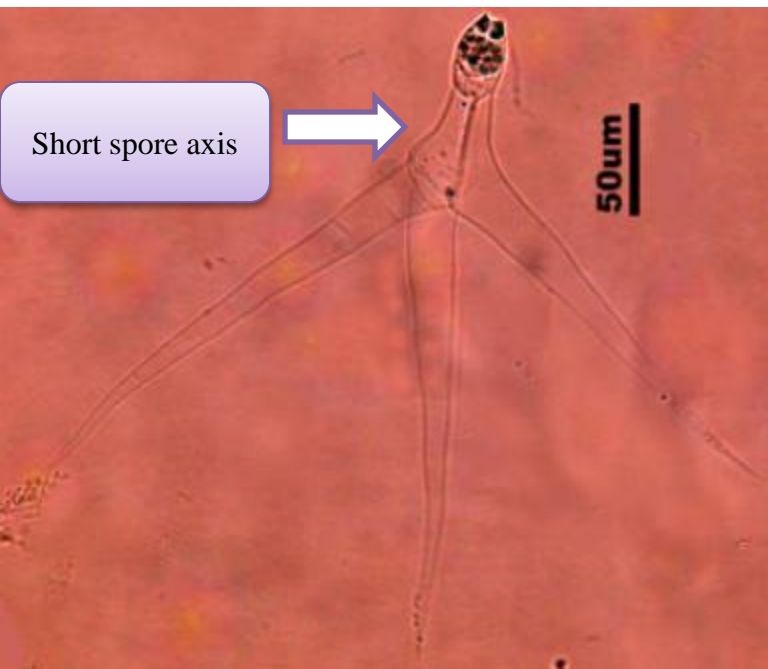
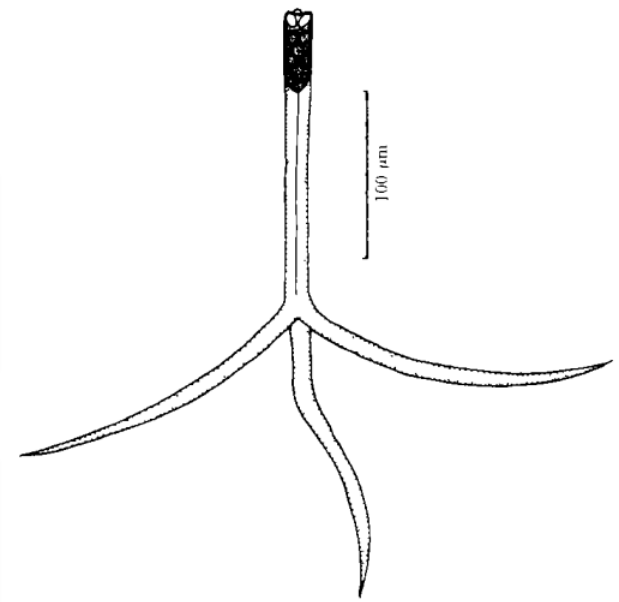


Phylogenetic relationship consistent with spore morphology (round spore body)

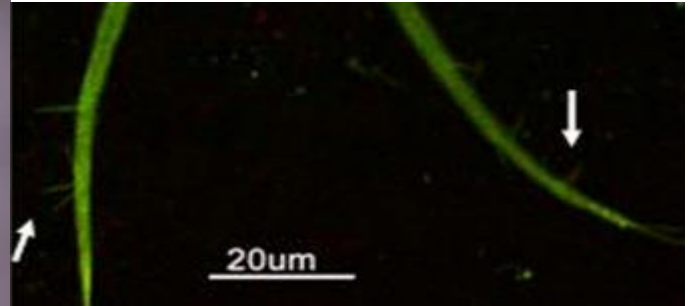
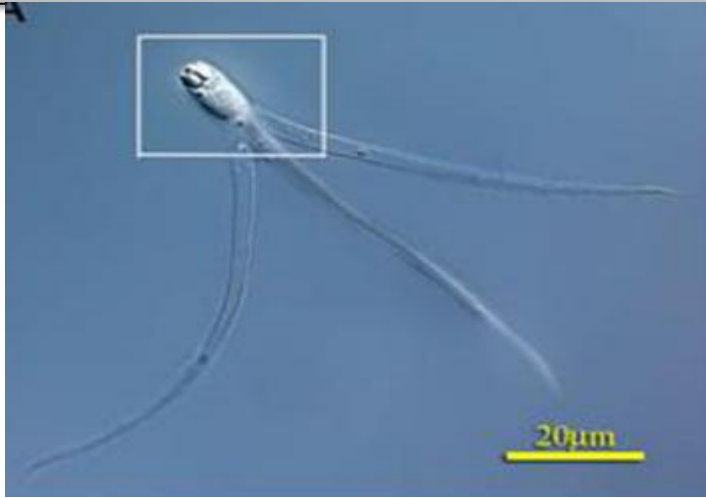
## Life cycle study in China

● Firstly reported *Triactinomyxon* type from *Branchiura sowerbyi* (Wang et al. 2000)  
no molecular data

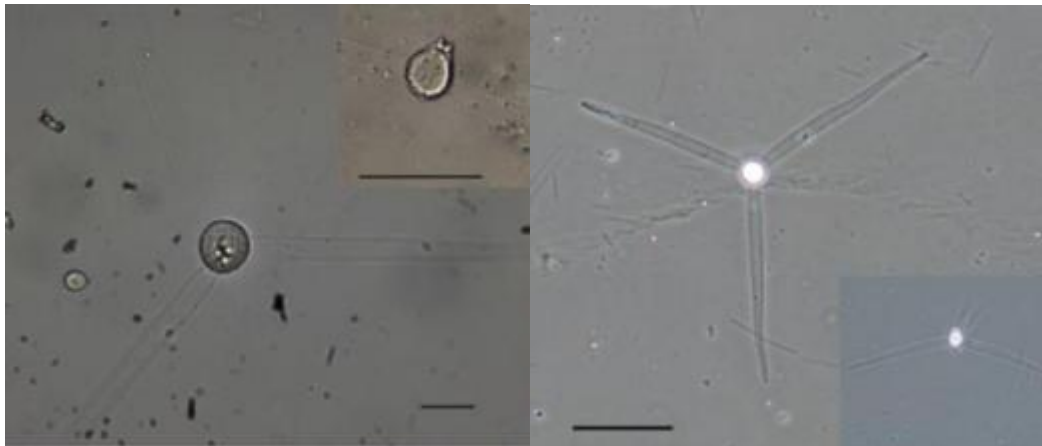
● Secondly *Triactinomyxon* type from *Branchiura sowerbyi* (Zhai et al. 2012)  
molecular data: HM107112



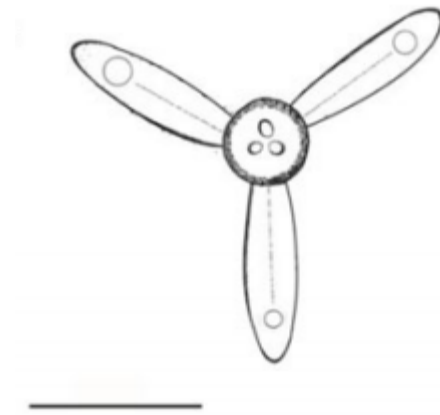
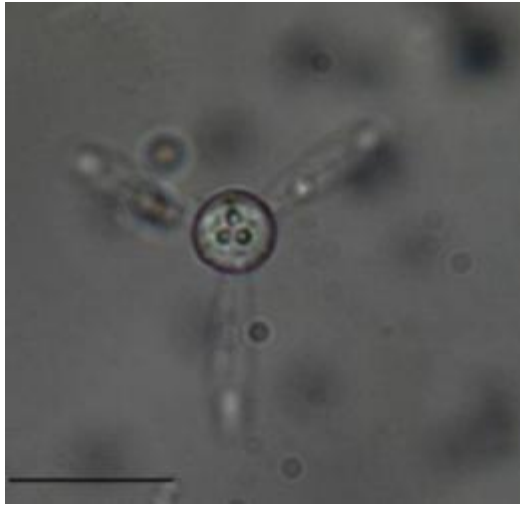
● Firstly *Raabeia* type from *Branchiura sowerbyi* (Zhai et al. 2013; Xi et al. 2013)  
molecular data: AB121146; HQ613409



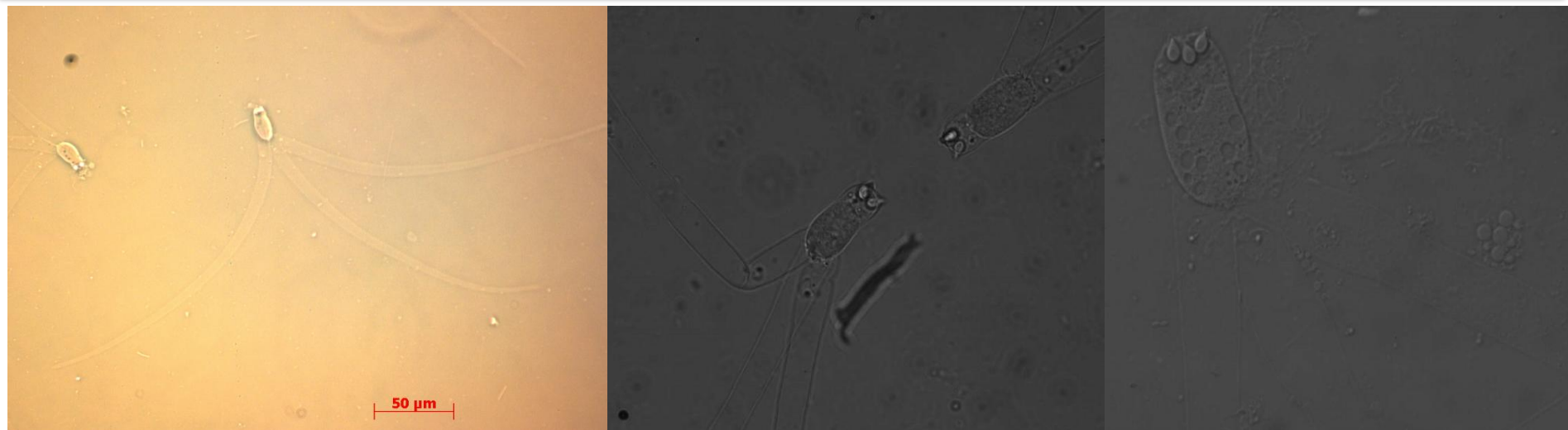
● Firstly *Aurantiactinomyxon* type from *Branchiura sowerbyi* (Xi et al. 2013); molecular HQ613406



● Firstly *Guyenotia* type from *Branchiura sowerbyi* (Xi et al. 2013); molecular data unavailable



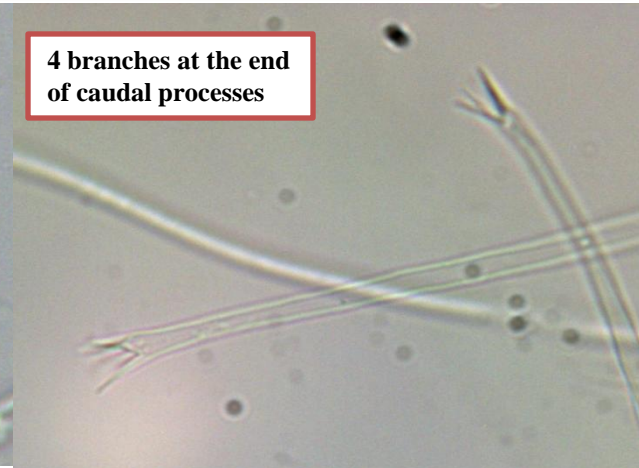
● Secondly *Raabeia* type from *Branchiura sowerbyi*  
molecular data: HQ613410



● Firstly *Echinactionmyxon* type from *Branchiura sowerbyi*;  
molecular data: EF690300



4 branches at the end  
of caudal processes



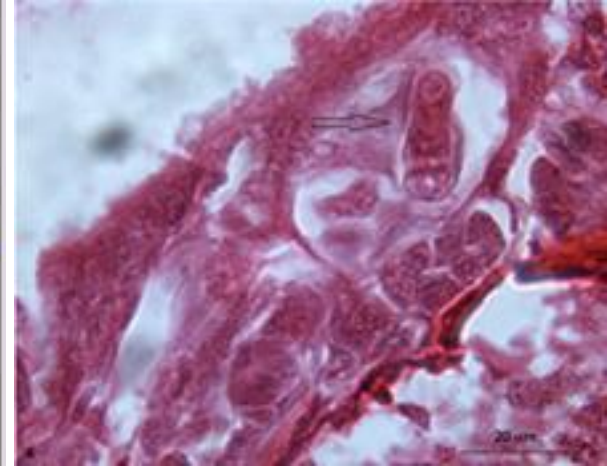
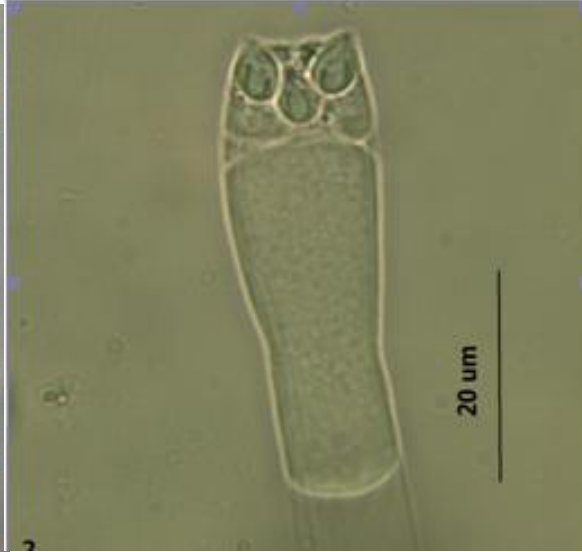
● Secondly *Echinactionmyxon* type from *Branchiura sowerbyi*;  
molecular data: unavailable



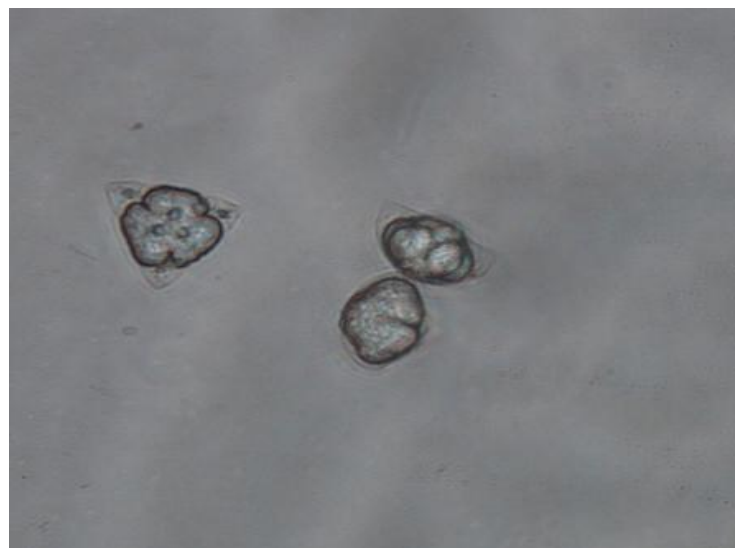
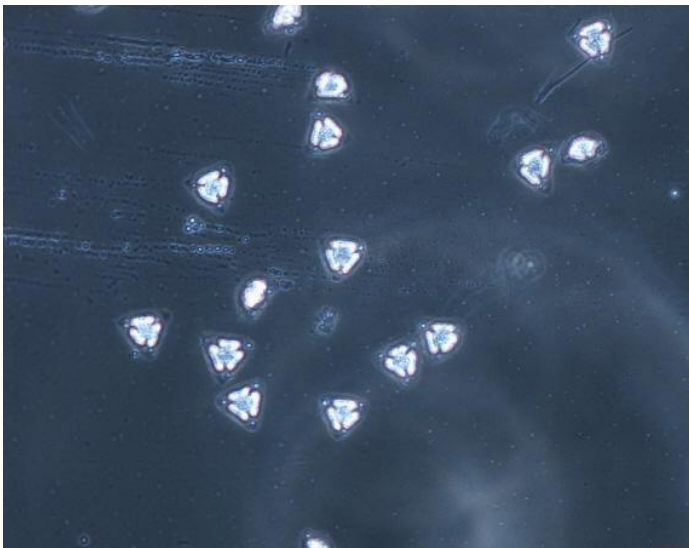
many branches at the  
end of caudal  
processes



● Thirdly *Triactinomyxon* from *Branchiura*  
molecular data: JX477771



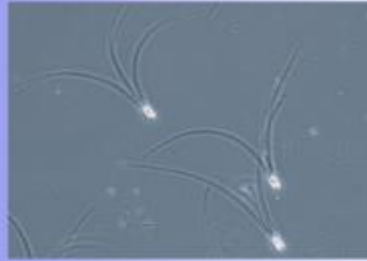
● Firstly *Neoactinomyxum* type from *Branchiura sowerbyi*;  
molecular data: KP642135, JX458816



*Myxobolus cultus*

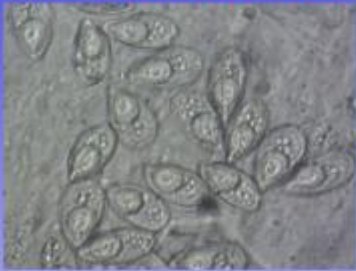


Raabiea

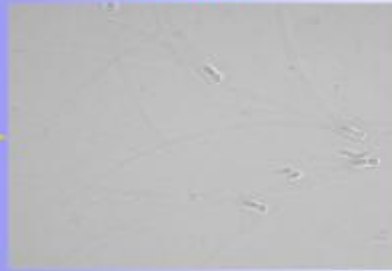


**Molecular data match**

*Thelohanellus wuhanensis*

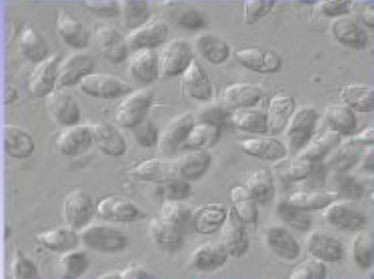


Raabeia



**Experimental infection  
and molecular match**

*Thelohanellus wangi*



Neoantinomyxum



**Molecular data match**

*Myxobolus wulii*



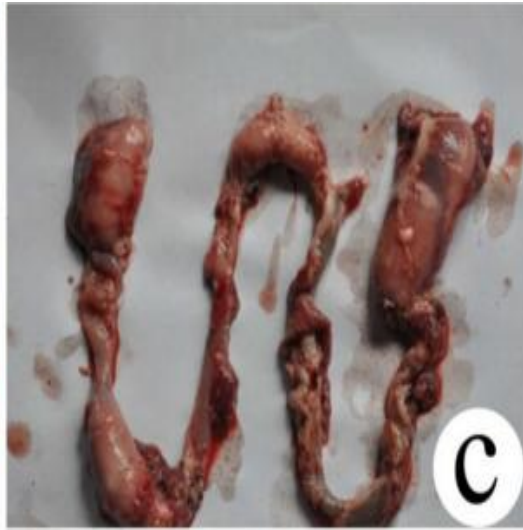
Echinactinomyxon



**Molecular data match**

# Current severe myxosporidiosis of common carp in China

Giant cystic diseases caused by *Thelohanellus kitauei*



Infection prevalence: 70%;

Mortality rate: up to 90%, especially Northern China;

Infection season: from early August to late October, especially Autumn;

Economic losses: 50 million per year;

Control strategy: no practical way

# Current severe myxosporidiosis of gibel carp in China

## 1. Pharynx myxosporidiosis caused by *Myxobolus honghensis*



Infection prevalence: 70%;

Mortality rate: up to 100%, especially in Northern Jiangsu Province;

Infection season: from early May to late October, especially summer;

Economic losses: 2 billion per year;

Control strategy: blocking the transmission and inactivating the infective actinospores by Ino-org;

## 2. Gill myxosporidiosis caused by *Thelohanellus wangi*



Infection prevalence: 60%;

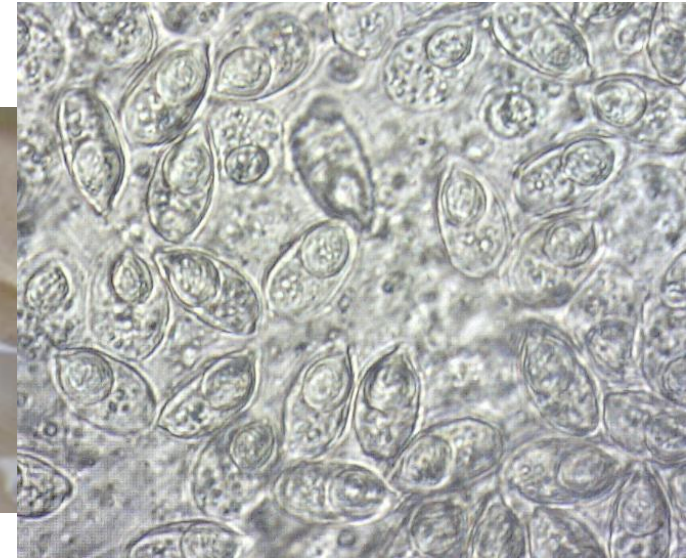
Mortality rate: up to 100%, especially under anoxia, crowded or other stress;

Infection season: only from early April to late June on fry fish below 7cm in body length;

Economic losses: 10 million per year;

Control strategy: blocking the transmission and inactivating the infective actinospores by Ino-org;

### 3.Skin myxosporidiosis caused by *Thelohanellus wuhanensis*



Infection prevalence: 70%;

Mortality rate: no directly lethal, but up to 100% under anoxia, crowded or other stress condition;

Infection season: only from early April to late June on fry fish below 7cm in body length;

Economic losses: 10 million per year;

Control strategy: blocking the transmission and inactivating the infective actinospores by Ino-org;

# Acknowledge

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Thank you for your attention